



Europäisches Patentamt
European Patent Office
Office européen des brevets



Publication number: **0 644 262 A2**

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: **94303401.7**

(22) Date of filing: **11.05.94**

(51) Int. Cl.⁶ **C12N 15/31, C07K 14/39,
C07K 14/37, C07K 16/14,
C12Q 1/68, G01N 33/569,
C12N 15/11**

(30) Priority: **24.05.93 JP 142523/93
28.12.93 JP 348893/93**

(43) Date of publication of application:
22.03.95 Bulletin 95/12

(34) Designated Contracting States:
CH DE ES FR GB IT LI

(71) Applicant: **TAKARA SHUZO CO. LTD.**
**609 Takenaka-cho
Fushimi-ku,
Kyoto-shi,
Kyoto-fu (JP)**

(72) Inventor: **Takashi, Okado**
**4-36-312, Honmaru-cho
Otsu-shi,
Shiga-ken (JP)**
Inventor: **Kazutoh, Takesako**
**4-20-208, Akibadai
Otsu-shi,
Shiga-ken (JP)**
Inventor: **Ikunoshin, Kato**
**1-1-150, Nanryo-cho
Uji-Shi,
Kyoto-fu (JP)**

(74) Representative: **Wakerley, Helen Rachael**
**Reddie & Grose,
16 Theobalds Road
London WC1X 8PL (GB)**

(54) **Gene coding for a protein regulating aureobasidin sensitivity.**

(57) An isolated gene coding for a protein regulating aureobasidin sensitivity. A process for cloning the gene with the use of the gene or a part of the same as a probe. A nucleic acid probe being hybridizable with the gene. An antisense DNA or RNA of the gene. A recombinant or transformant having the gene contained therein. An isolated protein regulating aureobasidin sensitivity and a process for producing the same by using the transformant. An antibody for the protein. A process for detecting the protein or the gene. A process for screening an antimycotic by using the protein or the transformant. This invention is useful in the diagnosis and treatment of diseases including mycoses.

EP 0 644 262 A2

This invention relates to a protein regulating the sensitivity to an antimycotic aureobasidin and a gene coding for this protein, namely, a gene coding for a protein regulating aureobasidin sensitivity. The present invention further relates to a series of the uses of the protein and the gene. Furthermore, it relates to an antibody against this protein and the use of the same.

Systemic mycoses including candidiasis have increased with an increase in immunocompromised patients in recent years due to, for example, the extended use of immunosuppressive drugs and acquired immunodeficiency syndrome (AIDS), and as opportunistic infection due to microbial substitution caused by the frequent use of widespectrum antibacterial antibiotics. Although drugs for treating mycoses such as amphotericin B, flucytosine and azole drugs (for example, fluconazole and miconazole) are now used to cope with this situation, none of them can achieve a satisfactory effect. Also, known diagnostic drugs are insufficient. For candidiasis, in particular, although there have been known several diagnostic drugs (for example, CAND-TEC for detection of candida antigen and LABOFIT for detection of D-arabinitol), none of them gives any satisfactory results in specificity or sensitivity.

The reasons for the delay in the development of remedies and diagnostic drugs for mycoses as described above are that fungi causing the mycoses are eukaryotic organisms similar to the host (i.e., man) and thus not largely different from man and that knowledges of fungi, in particular, pathogenic fungi are insufficient. Therefore it is difficult to distinguish fungi from man or to selectively kill fungi, which is responsible for the delay in the development of drugs for mycoses.

Recently the application of genetic engineering techniques such as antisense or PCR to the treatment and diagnosis of mycoses has been expected. However known genes which are applicable thereto and or proteins coded for by these genes are rare (PCT Pamphlet W092 03455). Regarding pathogenic fungi, there have been cloned in recent years an acid protease gene, which has been assumed to participate in the pathogenicity of Candida albicans (hereinafter referred to simply as C. albicans) and Candida tropicalis - (hereinafter referred to as C. tropicalis) causing candidiasis [B. Hube et al., J. Med. Vet. Mycol., 29, 129 - 132 (1991); Japanese Patent Laid-Open No. 49476 1993; and G. Togni et al., FEBS Letters, 286, 181 - 185 (1991)], a calmodulin gene of C. albicans [S.M. Saporito et al., Gene, 106, 43 - 49 (1991)] and a glycolytic pathway enzyme enolase gene of C. albicans [P. Sundstrom et al., J. Bacteriology, 174, 6789 - 6799 (1991)]. However, each of these genes and proteins coded for thereby is either indistinguishable from nonpathogenic fungi and eukaryotic organisms other than fungi or, if distinguishable therefrom, cannot serve as a definite action point for exhibiting any selective toxicity. Aureobasidin [Japanese Patent Laid-Open No. 138296 1990, No. 22995 1991, No. 220199 1991 and No. 279384 1993, Japanese Patent Application No. 303177 1992, J. Antibiotics, 44 (9), 919 - 924, ibid., 44 (9), 925 - 933, ibid., 44 (11), 1187 - 1198 (1991)] is a cyclic depsipeptide obtained as a fermentation product of a strain Aureobasidium pullulans No. R106. It is completely different in structure from other antimycotics. As Tables 1 and 2 show, aureobasidin A, which is a typical aureobasidin compound, exerts a potent antimycotic activity on various yeasts of the genus Candida including C. albicans which is a pathogenic fungus, Cryptococcus neoformans, Histoplasma capsulatum, Blastomyces dermatitidis and fungi of the genus Aspergillus (Japanese Patent Laid-open No. 138296 1990) but has an extremely low toxicity in mammal. Thus this compound is expected to be useful as an antimycotic being excellent in selective toxicity.

Hereinafter, Candida, Cryptococcus and Aspergillus will be abbreviated respectively as C., Cr. and A.

Table 1

Test strain	TIMM No.	MIC(μ g/ml)
<u>C. albicans</u>	0136	≤ 0.04
<u>C. albicans var. stellatoidea</u>	1308	≤ 0.04
<u>C. tropicalis</u>	0312	0.08
<u>C. kefyr</u>	0298	0.16
<u>C. parapsilosis</u>	0287	0.16
<u>C. krusei</u>	0270	≤ 0.04
<u>C. guilliermondii</u>	0257	0.08
<u>C. glabrata</u>	1062	≤ 0.04
<u>Cr. neoformans</u>	0354	0.63
<u>Cr. terreus</u>	0424	0.31
<u>Rhodotorula rubra</u>	0923	0.63
<u>A. fumigatus</u>	0063	20
<u>A. clavatus</u>	0056	0.16

Table 2

Test strain	TIMM No.	MIC(μ g/ml)
<u>A. nidulans</u>	0112	0.16
<u>A. terreus</u>	0120	5
<u>Penicillium commune</u>	1331	1.25
<u>Trichophyton mentagrophytes</u>	1189	10
<u>Epidermophyton floccosum</u>	0431	2.5
<u>Fonsecaea pedrosoi</u>	0482	0.31
<u>Exophiala werneckii</u>	1334	1.25
<u>Cladosporium bantianum</u>	0343	0.63
<u>Histoplasma capsulatum</u>	0713	0.16
<u>Paracoccidioides brasiliensis</u>	0880	0.31
<u>Geotrichum candidum</u>	0694	0.63
<u>Blastomyces dermatitidis</u>	0126	0.31

[Problems to be Solved by the Invention]

Each of the conventional antimycotics with a weak toxicity shows only a fungistatic effect, which has been regarded as a clinical problem. In contrast, aureobasidin has a fungicidal effect. From this point of view, it has been urgently required to clarify the mechanism of the selective toxicity to fungi of aureobasidin. However this mechanism still remains unknown.

Under these circumstances, the present invention aims at finding a novel protein regulating aureobasidin sensitivity through the clarification of the mechanism of the selective toxicity to fungi of aureobasidin. Accordingly, the present invention aims at finding a gene coding for a protein regulating aureobasidin sensitivity, providing a process for cloning this gene and the protein regulating aureobasidin sensitivity which is encoded by this gene, further providing an antisense DNA and an antisense RNA of this gene, providing a nucleic acid probe being hybridizable with this gene, providing a process for detecting this gene with the use of the nucleic acid probe, providing a process for producing the protein regulating aureobasidin sensitivity by using this gene and providing an antibody against the protein regulating aureobasidin sensitivity, and a process for detecting the protein regulating aureobasidin sensitivity by using this antibody.

[Means for Solving the Problems]

The present invention may be summarized as follows. Namely, the first aspect of the present invention relates to an isolated gene coding for a protein regulating aureobasidin sensitivity, that is, a gene regulating aureobasidin sensitivity. The second aspect of the invention relates to a process for cloning a gene regulating aureobasidin sensitivity which is characterized by using the gene regulating aureobasidin sensitivity of the first aspect or a part thereof as a probe. The third aspect of the invention relates to a nucleic acid probe which is hybridizable with a gene regulating aureobasidin sensitivity and comprises a sequence consisting of 15 or more bases. The fourth aspect of the invention relates to an antisense DNA of a gene regulating aureobasidin sensitivity. The fifth aspect of the invention relates to an antisense RNA of a gene regulating aureobasidin sensitivity. The sixth aspect of the invention relates to a recombinant plasmid having a gene regulating aureobasidin sensitivity contained therein. The seventh aspect of the invention relates to transformant having the above-mentioned plasmid introduced therinto. The eighth aspect of the invention relates to a process for producing a protein regulating aureobasidin sensitivity by using the above-mentioned transformant. The ninth aspect of the invention relates to an isolated protein regulating aureobasidin sensitivity. The tenth aspect of the invention relates to an antibody against a protein regulating aureobasidin sensitivity. The eleventh aspect of the invention relates to a process for detecting a protein regulating aureobasidin sensitivity by using the above-mentioned antibody. The twelfth aspect of the invention relates to a process for detecting a gene regulating aureobasidin sensitivity by the hybridization which is characterized by using the nucleic acid probe of the third aspect of the present invention. The thirteenth aspect of the invention relates to a process for screening an antimycotic by using the above-mentioned transformant or a protein regulating aureobasidin sensitivity.

The present inventors have found out that fungi such as Schizosaccharomyces pombe (hereinafter referred to simply as Schizo. pombe) and Saccharomyces cerevisiae (hereinafter referred to simply as S. cerevisiae) and, further, mammalian cells such as mouse lymphoma EL-4 cells are sensitive to aureobasidin, as Table 3 shows.

Table 3

Test strain or cell	MIC(μ g. ml)
<u>Schizo. pombe</u>	0.08
<u>S. cerevisiae</u>	0.31
mouse lymphoma EL-4	10
mouse lymphoma L5178Y	100
NRK-49F	12.5

The present inventors have mutagenized a wild-type strain of Schizo. pombe or S. cerevisiae, sensitive to aureobasidin, to thereby give resistant mutants. We have further successfully isolated a gene capable of conferring aureobasidin resistance (a resistant gene) from these resistant mutants and another gene capable of imparting aureobasidin sensitivity (a sensitive gene) from the corresponding sensitive cells. Furthermore, We have disclosed the existence of a protein encoded by each of these genes. By culturing cells which have been transformed by introducing the above-mentioned gene, We have succeeded in the expression of this gene. Furthermore, We have successfully found out a novel gene regulating aureobasidin sensitivity from another fungus being sensitive to aureobasidin by using a DNA fragment of the above-mentioned gene as a probe. In addition, We have clarified that the gene regulating aureobasidin sensitivity is essentially required for the growth of the cells and found out that the detection of this gene or a protein which is a gene product thereof with an antibody enables the diagnosis of diseases caused by these cells, for example, mycoses induced by fungi, and that an antisense DNA or an antisense RNA, which inhibits the expression of the gene regulating aureobasidin sensitivity being characteristic to the cells, is usable as a remedy for diseases caused by these cells, for example, mycoses induced by fungi, thus completing the present invention.

That is to say, pathogenic fungi listed in Tables 1 and 2 and fungi and mammalian cells listed in Table 3, each having a sensitivity to aureobasidin, each carries a protein regulating aureobasidin sensitivity and a gene coding for this protein. The term "a protein regulating aureobasidin sensitivity" as used herein means a protein which is contained in an organism having a sensitivity to aureobasidin. This protein is required for the expression of the sensitivity or resistance to aureobasidin. As a matter of course, a protein having 35% or more homology with the above-mentioned protein and having a similar function is also a member of the

protein regulating aureobasidin sensitivity according to the present invention. Furthermore, proteins obtained by modifying these proteins by the genetic engineering procedure are members of the protein regulating aureobasidin sensitivity according to the present invention. A gene regulating aureobasidin sensitivity means a gene which codes for such a protein regulating aureobasidin sensitivity as those described above and involves both of sensitive genes and resistant genes.

The first aspect of the present invention relates to a gene regulating aureobasidin sensitivity. This gene can be isolated in the following manner. First, aureobasidin sensitive cells (a wild-type strain) is mutagenized to thereby induce a resistant strain. From chromosome DNA or cDNA of this resistant strain, a DNA library is prepared and a gene capable of conferring a resistance (a resistant gene) is cloned from this library. Then a DNA library of a wild strain is prepared and a DNA molecule being hybridizable with the resistant gene is isolated from this library and cloned. Thus a sensitive gene can be isolated.

The mutagenesis is performed by, for example, treating with a chemical such as ethylmethane sulfonate (EMS) or N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) or by ultraviolet or other radiation. The cell that has acquired the resistance can be screened by culturing the mutagenized cells in a nutritional medium containing aureobasidin at an appropriate concentration under appropriate conditions. The resistant strain thus obtained may vary depending on the method and conditions selected for the mutagenesis. Also, strains differing in the extent of resistance from each other can be separated by changing the aureobasidin concentration or a temperature-sensitive resistant strain can be isolated by changing the temperature in the step of screening. There are a number of mechanisms of resistance to aureobasidin. Accordingly, a number of resistant genes can be isolated by genetically classifying these various resistant strains. In the case of a yeast, the classification may be performed by the complementation test. Namely, resistant strains are prepared from haploid cells. Next, diploid cells can be obtained by crossing resistant strains differing in mating type from each other. Then spores formed from these diploids are examined by the tetrad analysis.

As typical examples of the genes regulating aureobasidin sensitivity (named *aur*) according to the present invention, *aur1* and *aur2* genes may be cited. Typical examples of the *aur1* gene include *spaur1* gene isolated from *Schizo. pombe* and *scaur1* gene isolated from *S. cerevisiae*, while typical examples of the *aur2* gene include *scaur2* gene isolated from *S. cerevisiae*. Now, resistant genes (*spaur1^R*, *scaur1^R* and *scaur2^R*) isolated from resistant mutants by the present inventors and sensitive genes (*spaur1^S*, *scaur1^S* and *scaur2^S*) isolated from sensitive wild-type strains will be described.

Fig. 1 shows a restriction enzyme map of the genes *spaur1^R* and *spaur1^S* regulating aureobasidin sensitivity, Fig. 2 shows a restriction enzyme map of *scaur1^R* and *scaur1^S* and Fig. 3 shows a restriction enzyme map of *scaur2^R* and *scaur2^S*.

Schizo. pombe, which is sensitive to aureobasidin, is mutagenized with EMS and a genomic library of the resistant strain thus obtained is prepared. From this library, a DNA fragment containing a resistant gene (*spaur1^R*) and having the restriction enzyme map of Fig. 1 is isolated. This gene has a nucleotide sequence represented by SEQ ID No. 1 in Sequence Listing. The amino acid sequence of a protein encoded by this gene, which is estimated on the basis of this nucleotide sequence, is the one represented by SEQ ID No. 2 in Sequence Listing. By the hybridization with the use of this resistant gene as a probe, a DNA fragment containing a sensitive gene (*spaur1^S*) and having the restriction enzyme map of Fig. 1 is isolated from a sensitive strain. This gene has a nucleotide sequence represented by SEQ ID No. 3 in Sequence Listing. The amino acid sequence of a protein encoded by this gene, which is estimated on the basis of this nucleotide sequence, is the one represented by SEQ ID No. 4 in Sequence Listing. A comparison between the sequences of SEQ ID No. 3 and SEQ ID No. 1 reveals that a mutation from G to T occurs at the base at the position 1053, while a comparison between the sequences of SEQ ID No. 4 and SEQ ID No. 2 reveals that glycine at the residue 240 is converted into cysteine at the amino acid level, thus giving rise to the resistance.

Also, *S. cerevisiae*, which is sensitive to aureobasidin, is mutagenized with EMS and genomic libraries of two resistant strains thus obtained are prepared. From one of these libraries, a DNA fragment containing a resistant gene (*scaur1^R*) as a dominant mutant and having the restriction enzyme map of Fig. 2 is isolated, while a DNA fragment containing a resistant gene (*scaur2^R*) and having the restriction enzyme map of Fig. 3 is isolated from another library.

The nucleotide sequence of the coding region for the protein of the *scaur1^R* gene is the one represented by SEQ ID No. 5 in Sequence Listing. The amino acid sequence of the protein encoded by this gene, which is estimated on the basis of the above nucleotide sequence, is the one represented by SEQ ID No. 6 in Sequence Listing. By the hybridization with the use of this resistant gene *scaur1^R* as a probe, a DNA fragment containing a sensitive gene (*scaur1^S*) and having the restriction enzyme map of Fig. 2 is isolated from a sensitive strain. This gene has a nucleotide sequence represented by SEQ ID No. 7 in Sequence Listing. The amino acid sequence of a protein encoded by this gene, which is estimated on the

basis of this nucleotide sequence, is the one represented by SEQ ID No. 8 in Sequence Listing. A comparison between the sequences of SEQ ID No. 7 and SEQ ID No. 5 reveals that a mutation from T to A occurs at the base at the position 852, while a comparison between the sequences of SEQ ID No. 8 and SEQ ID No. 6 reveals that phenylalanine at the residue 158 is converted into tyrosine at the amino acid level, thus giving rise to the resistance. The *spaur1* gene has a 58% homology with the *scaur1* gene at the amino acid level. Thus it is obvious that they are genes coding for proteins having similar functions to each other. When genes and proteins being homologous in sequence with the *spaur1* and *scaur1* genes and with the proteins encoded thereby are searched from a data base, none having a homology of 35% or above is detected. Accordingly, it is clear that these genes and the proteins encoded thereby are novel molecules which have never been known hitherto.

By the hybridization with the use of the DNA fragment of the resistant gene *scaur2^R* as a probe, a DNA fragment containing a sensitive gene (*scaur2^S*) and having the restriction enzyme map of Fig. 3 is isolated from a sensitive strain.

The nucleotide sequence of this sensitive gene is the one represented by SEQ ID No. 9 in Sequence Listing and the amino acid sequence of the protein encoded by this gene, which is estimated on the basis of this nucleotide sequence, is the one represented by SEQ ID No. 10 in Sequence Listing. As the result of the homology search with the *scaur2^S* gene and the protein encoded thereby, it has been found out that cystic fibrosis transmembrane conductance regulator (CFTR) of mammals alone has a homology as low as 31%. Compared with this CFTR, however, the part having a high homology is limited to the region around the domain of the nucleotide binding. It is therefore obvious that the protein encoded by the *scaur2^S* gene is a protein which is completely different from CFTR in function and has never been known hitherto.

In order to prove the importance of the *aur1* gene in the growth of cells, genes for disrupting the *aur1* as shown in Fig. 4 and Fig. 5, in which genes coding for orotidine-5'-phosphate decarboxylase (*ura4⁺* in the case of *Schizo. pombe*, while *URA3* in the case of *S. cerevisiae*) have been introduced midway in the *aur1* gene, are prepared. When these *aur1* disrupted genes are introduced into *Schizo. pombe* and *S. cerevisiae* respectively, the cells having the *aur1* disrupted genes cannot grow at all. Thus it has been revealed that these genes and the proteins encoded thereby are essentially required for the growth of the yeast cells.

As the above examples clearly show, a gene regulating aureobasidin sensitivity can be isolated by using a organism having sensitivity to aureobasidin as a starting material and by carrying out the cloning with the use of various mutagenesis methods and or screening methods depending on the organisms or the methods. Also, genes being hybridizable with the above-mentioned genes are involved in the scope of the first aspect of the present invention. A gene regulating aureobasidin sensitivity can be isolated by the following method. The genomic DNA library of an organism having sensitivity to aureobasidin is integrated into, for example, a high-expression vector of a yeast and transformed into the yeast. Then a clone having aureobasidin resistance is selected from the transformants and DNA is recovered from this clone. Thus the resistant gene can be obtained. As a matter of course, genes obtained by modifying some part of the gene regulating aureobasidin sensitivity thus obtained by some chemical or physical methods are involved in the scope of the first aspect of the present invention.

The second aspect of the present invention relates to a process for cloning a gene regulating aureobasidin sensitivity which is characterized by using the gene regulating aureobasidin sensitivity of the first aspect of the present invention or a part thereof as a probe. Namely, by screening by the hybridization method or the polymerase chain reaction (PCR) method with the use of a part (consisting of at least 15 oligonucleotides) or the whole of the gene as obtained above, a gene coding for a protein having a similar function can be isolated.

For example, a pair of primers of SEQ ID No. 11 and SEQ ID No. 12 in Sequence Listing are synthesized on the basis of the DNA nucleotide sequence of the *spaur1^R* gene represented by SEQ ID No. 1. Then PCR is performed by using cDNA of *C. albicans*, which is a pathogenic fungus, as a template with the use of the above-mentioned primers. The PCR is carried out and the PCR products are electrophoresed on an agarose gel and stained with ethidium bromide. In Fig. 6, the lanes 1, 2 and 3 show the results obtained by using cDNA of *C. albicans*, cDNA of *S. cerevisiae* and cDNA of *Schizo. pombe* as a template, respectively. As shown in Fig. 6, a certain DNA fragment is specifically amplified.

By screening the genomic DNA library of *C. albicans* with the use of this DNA fragment as a probe, a DNA molecule having a gene (*caaur1*), which has the same function as that of the *spaur1* and *scaur1* genes and having the restriction enzyme map of Fig. 7 is obtained. The nucleotide sequence of this *caaur1* gene is the one represented by SEQ ID No. 13 in Sequence Listing and the amino acid sequence of the protein encoded by this gene, which has been estimated on the basis of the above nucleotide sequence, is the one represented by SEQ ID No. 14 in Sequence Listing. It has a high homology with the proteins encoded by the *spaur1* and *scaur1* genes.

By screening the genomic DNA library of *C. albicans* with the use of a DNA fragment comprising the whole length or a part of the *scaur2^S* gene represented by SEQ ID No. 9 in Sequence Listing as a probe, a DNA fragment containing gene (*caaur2*), which has the same function as that of the *scaur2* gene, and having the restriction enzyme map of Fig. 8 is obtained. The nucleotide sequence of a part of this *caaur2* gene is the one represented by SEQ ID No. 15 in Sequence Listing and the amino acid sequence of the region encoded by this gene, which has been estimated on the basis of this nucleotide sequence, is the one represented by SEQ ID No. 16 in Sequence Listing. It has a high homology with the corresponding region of the protein encoded by the *scaur2* gene.

The third aspect of the present invention relates to an oligonucleotide comprising 15 or more bases which serves as the above-mentioned nucleic acid probe and is hybridizable with the gene regulating aureobasidin sensitivity, for example, the DNA fragment having the restriction enzyme map as shown in Fig. 1, Fig. 2 or Fig. 3. This nucleic acid probe is usable in, for example, the hybridization in situ, the identification of a tissue wherein the above-mentioned gene can be expressed, and the confirmation of the presence of a gene or mRNA in various vital tissues. This nucleic acid probe can be prepared by ligating the above-mentioned gene or a gene fragment to an appropriate vector, introducing it into a bacterium, allowing it to replicate in the bacterium, extracting from a disrupted cell suspension, cleaving with a restriction enzyme capable of recognizing the vector-ligating site, electrophoresing and then excising from the gel. Alternatively, this nucleic acid probe can be constructed by the chemical synthesis with the use of a DNA synthesizer or gene amplification techniques by PCR on the basis of the nucleotide sequence of SEQ ID Nos. 1, 3, 5, 7, 9, 13, 15 or 21 in Sequence Listing. This nucleic acid probe can be labeled with a radioisotope or a fluorescent substance to thereby elevate the detection sensitivity at the use.

The fourth aspect of the present invention relates to an antisense DNA of the above-mentioned gene regulating aureobasidin sensitivity, while the fifth aspect of the present invention relates to an antisense RNA thereof. The introduction of the antisense DNA or antisense RNA into cells makes it possible to control the expression of the gene regulating aureobasidin sensitivity.

As examples of the antisense DNA to be introduced, antisense DNAs corresponding to the genes regulating aureobasidin sensitivity of SEQ ID Nos. 1, 3, 5, 7, 9, 13, 15 or 21 in Sequence Listing and some parts thereof may be cited. SEQ ID No. 17 in Sequence Listing shows an example of this antisense DNA. It represents the sequence of an antisense DNA of the gene regulating aureobasidin sensitivity of SEQ ID No. 1 in Sequence Listing. A fragment obtained by appropriately cleaving some part of such an antisense DNA, and a DNA synthesized depending on such an antisense DNA sequence may be used as the antisense DNA.

As examples of the antisense RNA to be introduced, antisense RNAs corresponding to the genes regulating aureobasidin sensitivity of SEQ ID Nos. 1, 3, 5, 7, 9, 13, 15 or 21 in Sequence Listing and some parts thereof may be cited. SEQ ID No. 18 in Sequence Listing shows an example of this antisense RNA. It represents the sequence of an antisense RNA of the gene regulating aureobasidin sensitivity of SEQ ID No. 1 in Sequence Listing. A fragment obtained by appropriately cleaving some part of such an antisense RNA, an RNA synthesized depending on such an antisense RNA sequence, and an RNA prepared with RNA polymerase in an *in vitro* transcription system by using the DNA corresponding to the gene regulating aureobasidin sensitivity of SEQ ID No. 1 or SEQ ID No. 3 in Sequence Listing or a part thereof may be used as the antisense RNA.

These antisense DNA and antisense RNA may be chemically modified so as to prevent degradation *in vivo* or to facilitate passage through a cell membrane. A substance capable of inactivating mRNA, for example, ribozyme may be linked thereto. The antisense DNA and antisense RNA thus prepared are usable in the treatment of various diseases such as mycoses accompanied by an increase in the amount of mRNA coding for a protein regulating aureobasidin sensitivity.

The sixth aspect of the present invention relates to a recombinant plasmid having a gene coding for a protein regulating aureobasidin sensitivity being integrated into an appropriate vector. For example, a plasmid, in which a gene regulating aureobasidin sensitivity gene has been integrated into an appropriate yeast vector, is highly useful as a selection marker gene, since a transformant can be easily selected thereby with the guidance of the chemical resistance by using aureobasidin.

Also, the recombinant plasmid can be stably carried by, for example, *Escherichia coli*. Examples of vectors which are usable in this case include pUC118, pWH5, pAU-PS, TraplexII9 and pTV118. pAU-PS having the *spaur1^S* gene integrated therein is named pSPAR1. pWH5 having the *spaur1^S* gene integrated therein is named pSCAR1. pWH5 having the *scaur2^R* gene integrated therein is named pSCAR1. TraplexII9 vector having the *caaur1* gene integrated therein is named pCAAR1. pTV118 vector having a part of the *caaur2* gene integrated therein is named pCAAR2N. Each of these recombinant plasmids is transformed into *E. coli*. It is also possible to express these plasmids in an appropriate host. Such a gene is reduced

exclusively into the open reading frame (ORF) to be translated into a protein by cleaving with an appropriate restriction enzyme, if necessary, and then bound to an appropriate vector. Thus an expression recombinant plasmid can be obtained. When *E. coli* is used as the host, plasmids such as pTV118 may be used as a vector for the expression plasmid. When a yeast is used as the host, plasmids such as pYES2 may be used as the vector. When mammalian cells are used as the host, plasmids such as pMAMneo may be used as the vector.

The seventh aspect of the present invention relates to a transformant having the above-mentioned recombinant plasmid which has been introduced into an appropriate host. As the host, *E. coli*, yeasts and mammalian cells are usable. *E. coli* JM109 transformed by pSPAR1 having the spaur1^S gene integrated therein has been named and designated as *Escherichia coli* JM109 pSPAR1 and deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1 chome Tsukuba-shi Ibaraki-ken 305, JAPAN), in accordance with the Budapest Treaty under the accession number FERM BP-4485. *E. coli* HB101 transformed by pSCAR1 having the scaur1^S gene integrated therein has been named and designated as *Escherichia coli* HB101 pSCAR1 and deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4483. *E. coli* HB101 transformed by pSCAR2 having the scaur2^R gene integrated therein has been named and designated as *Escherichia coli* HB101 pSCAR2 and deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4484. *E. coli* HB101 transformed by pCCAR1 having the caaur1^S gene integrated therein has been named and designated as *Escherichia coli* HB101 pCAAR1 and deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4482. *E. coli* HB101 transformed by pCAAR2N having a part of the caaur2 gene integrated therein has been named and designated as *Escherichia coli* HB101 pCAAR2N and deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4481.

A transformant capable of expressing a protein regulating aureobasidin sensitivity can be obtained by transforming an expression recombinant plasmid into an appropriate host, as described above. For example, a yeast having a recombinant plasmid as shown in Fig. 9 introduced thereto is usable for this purpose.

The eighth aspect of the present invention relates to a process for producing a protein regulating aureobasidin sensitivity which comprises incubating a transformant according to the sixth aspect of the present invention, which contains a gene coding for this protein, in an appropriate nutritional medium, allowing the expression of the protein, then recovering the protein from the cells or the medium and purifying the same. For the expression of the gene coding for this protein, *E. coli*, a yeast or mammalian cells are employed as a host. When the yeast having the recombinant plasmid of Fig. 9 is incubated in a medium containing galactose, for example, the protein regulating aureobasidin sensitivity which is encoded by the scaur1^S gene can be expressed.

The ninth aspect of the present invention relates to an isolated protein regulating aureobasidin sensitivity. As examples of such a protein, those encoded by the above-mentioned spaur1, scaur1, scaur2, caaur1 and caaur2 genes can be cited.

The spaur1^S gene codes for a protein having an amino acid sequence represented by SEQ ID No. 4 in Sequence Listing, while the scaur1^S gene codes for a protein having an amino acid sequence represented by SEQ ID No. 8 in Sequence Listing. By the northern hybridization with the use of a DNA fragment of the spaur1 gene as a probe, mRNAs are detected from a sensitive strain (Fig. 10). Thus the expression of the spaur1 gene is confirmed.

Fig. 10 is an autoradiogram showing the results of the northern hybridization wherein mRNAs obtained from cells of a sensitive strain of *Schizo. pombe* in the logarithmic growth phase (lane 1), cells of a resistant strain in the logarithmic growth phase (lane 2), cells of the sensitive strain in the stationary phase (lane 3) and cells of the resistant strain in the stationary phase (lane 4) are electrophoresed on a 1.2% agarose gel containing formaldehyde.

The tenth aspect of the present invention relates to an antibody against the above-mentioned protein regulating aureobasidin sensitivity. For example proteins having amino acid sequences of SEQ ID Nos. 2, 4, 6, 8, 10, 14, 16 or 22 in Sequence Listing and peptides comprising some parts of these amino acid sequences may be used as an antigen. The former antigens can be prepared through the expression in a transformant followed by purification, while the latter antigens can be synthesized on, for example, a marketed synthesizer. The antibody is produced by the conventional method. For example, an animal such as a rabbit is immunized with the above-mentioned protein or a peptide fragment together with an adjuvant

to thereby give a polyclonal antibody. A monoclonal antibody can be produced by fusing antibody-producing B cells, which have been obtained by immunizing with an antigen, with myeloma cells, screening hybridomas producing the target antibody, and incubating these cells. As will be described hereinafter, these antibodies are usable in the treatment and diagnosis for animal and human diseases in which the above-mentioned proteins participate, such as mycoses.

For example, a peptide corresponding to the part of the 103- to 113-positions in the amino acid sequence of SEQ ID No. 8 is synthesized on a synthesizer and then bound to a carrier protein. Then a rabbit is immunized therewith and thus a polyclonal antibody is obtained. In the present invention, keyhole limpet hemocyanin (KLH) is used as the carrier protein. Alternatively, bovine serum albumin and ovalbumin are usable therefor.

The eleventh aspect of the present invention relates to a process for detecting a protein regulating aureobasidin sensitivity by using the above-mentioned antibody. The detection can be carried out by detecting the binding of the antibody to the protein or measuring the amount of binding. For example, the protein or the cells producing the same can be detected by treating with a fluorescence-labeled antibody and then observing under a fluorescence microscope. The amount of the antibody bound to the protein can be measured by various known methods. For example, *S. cerevisiae* cells are stained by the immunofluorescent antibody technique by using the above-mentioned antibody and a secondary antibody such as FITC-labeled antirabbit antibody. Thus it is clarified that the protein encoded by the *scaur1* gene is distributed all over the cells. Further, a yeast having the recombinant plasmid of Fig. 9 introduced thereto is incubated in a medium containing glucose or galactose. The cells thus obtained are disrupted with glass beads and proteins are solubilized. Then these proteins are separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and the western blotting is carried out in the conventional manner by using the above-mentioned polyclonal antibody and peroxidase-labeled anti-rabbit antibody. Consequently, the protein encoded by the *scaur1* gene can be detected, as Fig. 11 shows.

Fig. 11 shows the results of the western blotting wherein the proteins prepared from the cells obtained by the incubation in the presence of glucose (lane 1) or galactose (lane 2) are subjected to SDS-PAGE. A main band binding to the polyclonal antibody of the present invention is detected at around 38 kDa.

The twelfth aspect of the present invention relates to a process for detecting a gene regulating aureobasidin sensitivity, for example, mRNA at the expression of a protein, by using the above-mentioned oligonucleotide as a nucleic acid probe. This process is applicable to the diagnosis for various diseases, including mycoses, associated with an abnormal amount of mRNA coding for the protein. For example, nucleic acids are precipitated from disrupted cells and mRNA is hybridized with a radioisotope-labeled nucleic acid probe on a nitrocellulose membrane. The amount of binding can be measured by autoradiography (Fig. 10) or with a scintillation counter.

The thirteenth aspect of the present invention relates to a process for efficient screening of a novel antimycotic by using the transformant of the seventh aspect of the present invention or the protein regulating aureobasidin sensitivity of the ninth aspect of the present invention. For example, a drug exerting its effect on the protein or the gene of the present invention can be efficiently found out through a comparison of the activity on a transformant containing a sensitive gene with the activity on a transformant containing a resistant gene or a comparison between the activities on transformants differing in expression level from each other. Also, the screening can be efficiently carried out by measuring the affinity for the protein of the present invention, for example, the activity of inhibiting the binding of radiolabeled-aureobasidin to the protein.

[Brief Description of the Drawings]

[Fig. 1]

Restriction enzyme map of the genes *spaur1^R* and *spaur1^S* regulating aureobasidin sensitivity.

[Fig. 2]

Restriction enzyme map of *scaur1^R* and *scaur1^S*.

[Fig. 3]

Restriction enzyme map of *scaur2^R* and *scaur2^S*.

[Fig. 4]

Structure of a DNA for disrupting the Schizo. pombe spaur1^S gene.

[Fig. 5]

Structure of a DNA for disrupting the S. cerevisiae scaur1^S gene.

[Fig. 6]

Results of the detection of the aur1 gene caaur1 carried by C. albicans by the PCR method.

[Fig. 7]

Restriction enzyme map of the caaur1 gene carried by C. albicans.

[Fig. 8]

Restriction enzyme map of the caaur2 gene.

[Fig. 9]

Structure of a plasmid YEpSCARW3 for expressing the scaur1 gene.

[Fig. 10]

Results of the northern hybridization of the spaur1 gene of Schizo. pombe.

[Fig. 11]

Results of the detection of the scaur1 protein by using an antibody.

[Fig. 12]

Restriction enzyme map of pAR25.

[Examples]

To further illustrate the present invention in greater detail, the following Examples will be given. However it is to be understood that the present invention is not restricted thereto.

Example 1: Cloning of a gene regulating aureobasidin sensitivity originating in fission yeast Schizo. pombe

1-a) Separation of aureobasidin-resistant mutant of Schizo. pombe

About 1×10^8 cells of a Schizo. pombe haploid cell strain JY745 (mating type h⁻, genotype ade6-M210, leu1, ura4-D18) exhibiting a sensitivity to aureobasidin at a concentration of 0.08 µg/ml were suspended in 1 ml of a phosphate buffer containing 0.9% NaCl. Then the cells were mutagenized with EMS at a final concentration of 3% at 30 °C for 90 minutes. After neutralizing by adding 8 ml of 5% sodium thiosulfate, the cells thus treated were harvested by centrifugation (2500 r.p.m., 5 minutes), washed twice with 6 ml of physiological saline and then suspended in 2 ml of a YEL medium (3% of glucose, 0.5% of yeast extract). The suspension was incubated at 30 °C for 5 hours under stirring and then spread on a YEA plate (the YEL medium containing 1.5% of agar) containing 5 µg/ml of aureobasidin A. After incubating at 30 °C for 3 to 4 days, two or three aureobasidin-resistant colonies were formed per 1×10^8 cells. After carrying out the mutagenesis several times, five clone mutants, i.e., THR01, THR04, THR05, THR06 and THR07 were obtained. These mutants were resistant to more than 25 µg/ml of aureobasidin A but the same as the parent strain in the sensitivity to cycloheximide and amphotericin B. Therefore it is estimated that they are not mutants having a multiple drug resistance but ones having a resistance specific to

aureobasidin.

1-b) Genetic analysis

Each of the above-mentioned resistant strains THR01, THR04, THR05, THR06 and THR07 was crossed with normal cells of *Schizo. pombe* LH121 strain (mating type h⁺, genotype ade6-M216, ura4-D18) differing in mating type. Diploid cells obtained were examined about the resistance to aureobasidin. Similar to the resistant strains, the five diploids formed by crossing the resistant strains with the normal one were resistant to 25 µg/ml of aureobasidin A, thus proving that these resistant mutations were dominant. To perform the tetrad analysis, the above-mentioned diploids were subsequently inoculated on an MEA medium (3% of malt extract, 2.5% of agar) for sporulation and incubated at 25 °C for 2 days. Prior to the meiosis, the diploid cells replicated DNA on the MEA medium and then underwent the meiosis to form asci each containing four ascospores of the haploid. These spores were separated with a micromanipulator and allowed to germinate on the YEA plate, followed by the formation of colonies. Then the resistance to aureobasidin of these colonies was examined. Among four spores contained in an ascus, the separation of the sensitivity versus the resistance showed 2 : 2. This result indicates that the aureobasidin resistant mutation was induced by a mutation in single gene. Further, the complementation test was performed in order to confirm whether the resistant genes of the above-mentioned five mutants were identical with each other or not. For example, a mutant of the mating type h⁺, which had been obtained by crossing the mutant THR01 with the LH121 strain in the above tetrad analysis, was crossed with another variant THR04 (mating type h⁻) on the MEA plate as described above and, after sporulation, the tetrad analysis was carried out. As a result, all of the colonies formed from four ascospores showed resistance to aureobasidin, which indicates that the mutational genes of THR01 and THR04 are the same with each other. Similarly, the five mutants were examined and it was thus found out that all mutations occurred on the same gene. This gene regulating aureobasidin sensitivity is named spaur1, the normal gene (sensitive gene) is named spaur1^S and the mutational gene (resistant gene) is named spaur1^R.

1-c) Preparation of genomic library of aureobasidin resistant strain

Genomic DNA was extracted and purified from the aureobasidin resistant strain THR01 by the method of P. Philippsen et al. [Methods in Enzymology, 194, 169 - 175 (1991)]. The purified genomic DNA (8 µg) was partially digested by treating with 5 U of a restriction enzyme HindIII at 37 °C for 10 minutes, deproteinized with phenol/chloroform and precipitated with ethanol. The partially digested DNA was electrophoresed on a 0.8% agarose gel and DNA in the region of 3 to 15 kb was extracted. The DNA thus obtained was ligated with a yeast-*E. coli* shuttle vector pAU-PS (2 µg) which had been completely digested with HindIII by using a DNA ligation kit (manufactured by Takara Shuzo Co., Ltd.) and then transformed into *E. coli* HB101. Thus a genomic library of the aureobasidin resistant strain was formed. *E. coli* containing this genomic library was incubated in 50 ml of an LB medium (1% of bacto trypton, 0.5% of bacto yeast extract, 0.5% of sodium chloride) containing 100 µg/ml of ampicillin and 25 µg/ml of tetracycline at 37 °C overnight. Then a plasmid was recovered and purified from the *E. coli* cells.

1-d) Expression and cloning of aureobasidin resistant gene spaur1^R

The plasmid originating in the genomic library of the aureobasidin resistant strain as prepared above was transformed into a strain *Schizo. pombe* JY745 by the method of Okazaki et al. [Nucleic Acid Research, 18, 6485 - 6489 (1990)]. The transformed cells were spreaded on a minimum medium SD plate [0.67% of yeast nitrogen base without amino acids (manufactured by Difco), 2% of glucose, 2% of agar] containing 75 µg/ml of adenine sulfate and 50 µg/ml of leucine. After incubating at 30 °C for 3 to 4 days, the colonies thus formed were replicated onto an SD plate containing 5 µg/ml of aureobasidin A, 75 µg/ml of adenine sulfate and 50 µg/ml of leucine. It is conceivably that a colony propagated on this plate may have the plasmid containing the aureobasidin resistant gene. This colony was inoculated into 5 ml of a liquid SD medium containing 75 µg/ml of adenine sulfate and 50 µg/ml of leucine. After incubating at 30 °C for 2 days, the plasmid was recovered from the propagated cells by the method of I. Hagan et al. [J. Cell Sci., 91, 587 - 595 (1988)]. Namely, the cells were harvested from the culture (5 ml) by centrifugation and then suspended in 1.5 ml of 50 mM citrate phosphate buffer containing 1.2 M of sorbitol and 2 mg/ml of Zymolyase. Then the suspension was maintained at 37 °C for 60 minutes. The cells were collected by centrifuging at 3,000 r.p.m. for 30 seconds and suspended in 300 µl of a TE [10 mM of Tris-HCl, pH 8, 1 mM of EDTA] solution. After adding 35 µl of 10% SDS, the mixture was maintained at 65 °C for 5 minutes.

After adding 100 μ l of 5 M potassium acetate, the mixture was allowed to stand in ice for 30 minutes. Then it was centrifuged at 10,000 r.p.m. at 4°C for 10 minutes and a plasmid DNA was purified from the supernatant by using EASYTRAP™ (manufactured by Takara Shuzo Co., Ltd.).

This plasmid was transformed into *E. coli* HB101 and a plasmid DNA was prepared from *E. coli* colonies formed on an LB medium containing ampicillin and tetracycline. This plasmid, which contained a DNA of 4.5 kb, was named pAR25. Fig. 12 shows the restriction enzyme map of the DNA of 4.5 kb in pAR25. To specify the gene region, *Hind*III fragments or *Sac*I fragments of various sizes were subcloned into the pAU-PS vector. These DNAs were transformed into normal JY745 cells by the above-mentioned method of Okazaki et al. and the acquisition of aureobasidin resistance was examined. As a result, it is revealed that a *Hind*III-*Sac*I 2.4 kb DNA fragment contains the *spaur*1^R gene. The restriction enzyme map of this DNA segment containing the aureobasidin resistant gene *spaur*1^R is shown in Fig. 1. This fragment was cloned into a pUC118 vector (named pUARS2R) and then the DNA nucleotide sequence was identified (SEQ ID No. 1 in Sequence Listing). From this nucleotide sequence, it is revealed that the *spaur*1^R gene code for a protein having an amino acid sequence represented by SEQ ID No. 2 in Sequence Listing.

1-e) Cloning of aureobasidin sensitive gene *spaur*1^S

By the same method as the one employed in the above c), genomic DNA was extracted and purified from normal cells. After partially digesting with *Hind*III, a genomic library of the normal cells was constructed. An *E. coli* stock containing this library DNA was spread on an LB agar medium containing ampicillin and tetracycline and incubated overnight at 37°C. The colonies thus formed were transferred onto a nylon membrane (Hybond™-N, manufactured by Amersham) and the colony hybridization was performed.

As a probe, the above-mentioned DNA fragment (2.4 kb) obtained by cleaving the *spaur*1^R gene with *Hind*III-*Sac*I and labeled with [α -³²P] dCTP by using a random primer DNA labeling kit (manufactured by Takara Shuzo Co., Ltd.) was used. As the results of screening of 5×10^4 colonies, five clones being hybridizable with the probe were obtained. Plasmids were purified from *E. coli* cells of these five clones. As the result of the cleavage with restriction enzymes, it was found out that all of these clones contained the same DNA fragment of 4.5 kb (named pARN1). The restriction enzyme map of the DNA of 4.5 kb in pARN1 was identical with that of pAR25 shown in Fig. 10. Therefore, a *Hind*III-*Sac*I 2.4 kb DNA fragment which was a region containing the *spaur*1^S gene was prepared from pARN1. Then it was cloned into the pAU-PS vector and this plasmid was named pSPAR1.

By using this plasmid pSPAR1, a strain *E. coli* JM109 was transformed and the transformant thus obtained was named and designated as *Escherichia coli* JM109 pSPAR1. It has been deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4485. This DNA fragment containing the aureobasidin sensitive gene *spaur*1^S had the restriction enzyme map shown in Fig. 1 and the DNA nucleotide sequence thereof was the one represented by SEQ ID No. 3 in Sequence Listing. Based on this nucleotide sequence, it has been revealed that the *spaur*1^S gene codes for a protein having the amino acid sequence represented by SEQ ID No. 4 in Sequence Listing and, when compared with the resistant gene *spaur*1^R, the amino acid at the residue 240 has been changed from glycine into cysteine.

Example 2: Cloning of aureobasidin sensitive genes *scaur*1 and *scaur*2 originating in budding yeast *S. cerevisiae*

2-a) Separation of aureobasidin resistant mutant of *S. cerevisiae*

A strain *S. cerevisiae* DKD5D (mating type a, genotype *leu*2-3 112, *trp*1, *his*3) having a sensitivity to aureobasidin at a concentration of 0.31 μ g/ml was mutagenized with EMS in the same manner as the one employed in the case of *Schizo. pombe*. Then resistant mutants were separated on an agar plate of a complete nutritional medium YPD (1% of yeast extract, 2% of polypeptone, 2% of glucose) containing 5 μ g/ml or 1.5 μ g/ml of aureobasidin A. After repeating the mutagenesis several times, 34 mutant clones were obtained. These mutants were resistant to more than 25 μ g/ml of aureobasidin A and estimated as having not a multiple drug resistance mutation but a aureobasidin-specific resistance mutation.

2-b) Genetic analysis

Similar to the above-mentioned case of Schizo. pombe, the genetic analysis using the tetrad analysis and the complementation test was performed. As a result, the genes could be classified into two types. These genes regulating aureobasidin sensitivity were named *scaur1* and *scaur2*, the resistant genes isolated from the resistant mutant were named *scaur1^R* and *scaur2^R*, and the sensitive genes isolated from the sensitive wild-type strain were named *scaur1^S* and *scaur2^S*, respectively.

The R94A strain had a gene with dominant mutation (*scaur1^R*). It has been further clarified that the *scaur1* gene is located in the neighborhood of the *met14* gene of the eleventh chromosome.

2-c) Preparation of genomic library of aureobasidin resistant strain having aureobasidin resistant gene *scaur1^R*

Genomic DNA was extracted and purified from the aureobasidin resistant strain R94A by the above-mentioned method of P. Philippsen et al. The purified genomic DNA (8 μ g) was partially digested by treating with 5 U of a restriction enzyme *Hind*III at 37°C for 10 minutes, deproteinized with phenol-chloroform and precipitated with ethanol. The partially digested DNA thus obtained was electrophoresed on a 0.8% agarose gel and DNA in the region of 3 to 15 kb was extracted. The DNA thus obtained was ligated with a yeast-*E. coli* shuttle vector pWH5 (2 μ g) which had been completely digested with *Hind*III by using a DNA ligation kit and then transformed into *E. coli* HB101. Thus a genomic library was formed. *E. coli* containing this genomic library was cultured in 50 ml of an LB medium containing ampicillin and tetracycline at 37°C overnight. Then a plasmid was recovered and purified from the *E. coli* cells.

2-d) Expression and cloning of aureobasidin resistant gene *scaur1^R*

The above-mentioned genomic library of the R94A strain was transformed into *S. cerevisiae* SH3328 (mating type α , genotype *ura3-52, his4, thr4, leu2-3* • 112) in accordance with the method of R.H. Schiestl et al. [Current Genetics, 16, 339 - 346 (1989)]. The transformed cells were spread on a minimum medium SD plate [0.67% of yeast nitrogen base without amino acids, 2% of glucose, 2% of agar] containing 25 μ g/ml of uracil, 35 μ g/ml of histidine and 500 μ g/ml of threonine. After incubating at 30°C for 3 to 4 days, the colonies thus formed were replicated onto a YPD agar plate containing 1.5 μ g/ml of aureobasidin A. A colony thus formed was inoculated into 5 ml of a liquid YPD medium. After incubating at 30°C for 2 days, a plasmid DNA was recovered from the propagated cells by the above-mentioned method of I. Hagan et al. This plasmid was transformed into a yeast again and it was confirmed that the obtained transformant had acquired aureobasidin resistance. This plasmid, which contained a DNA of 3.5 kb, was named pWTCR3. Neither the DNA fragment of 2.0 kb nor the DNA fragment of 1.5 kb obtained by cleaving with *Hind*III exhibited any aureobasidin resistant activity alone. Thus it is confirmed that the gene is contained in the DNA fragment of 3.5 kb. Fig. 2 shows the restriction enzyme map of this DNA fragment of 3.5 kb containing the aureobasidin resistant gene *scaur1^R*. The *Hind*III fragments of 1.5 kb and 2 kb were each cloned into pUC118, followed by the determination of the DNA nucleotide sequence (SEQ ID No. 5 in Sequence Listing). From this nucleotide sequence, it has been revealed that the *scaur1^R* gene codes for a protein having an amino acid sequence represented by SEQ ID No. 6 in Sequence Listing.

2-e) Cloning of aureobasidin sensitive gene *scaur1^S* corresponding to aureobasidin resistant gene *scaur1^R*

By the same method as the one employed in the above Example 2-c), genomic DNA was extracted and purified from the parent strain *S. cerevisiae* DKD5D. After partially digesting with *Hind*III, the DNA was ligated with pWH5 and transformed into *E. coli* HB101. Thus a genomic library of the normal cells was formed. An *E. coli* stock containing this library DNA was spread on an LB agar medium containing ampicillin and tetracycline and incubated overnight at 37°C. The colonies thus formed were transferred onto a nylon membrane (Hybond™-N) and the colony hybridization was carried out. As a probe, the DNA fragment of 3.5 kb obtained in the above Example 2-d) and labeled with [α -³²P] dCTP by using a random primer DNA labeling kit (manufactured by Takara Shuzo Co., Ltd.) was used. As the results of screening of 2 x 10⁴ colonies, seven clones being hybridizable with the probe were obtained. Plasmids were purified from *E. coli* cells of these clones. As the result of the cleavage with restriction enzymes, one of these clones contained a DNA fragment of 3.5 kb. This DNA fragment had the restriction enzyme map of Fig. 2 and thus judged as containing the *scaur1^S* gene. The plasmid containing this DNA fragment was named pSCAR1, while *E. coli* HB101 having this plasmid introduced therein was named and designated as *Escherichia coli*

HB101 pSCAR1. This strain has been deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4483. The DNA fragment of 3.5 kb obtained by partially digesting pSCAR1 with HindIII was subcloned into pUC118 and the nucleotide sequence thereof was determined (SEQ ID No. 7 in Sequence Listing). A comparison with the resistant gene indicates that the base at the position 852 has been changed from T into A and, due to this replacement, the amino acid has been converted from phenylalanine into tyrosine (SEQ ID No. 8 in Sequence Listing).

2-f) Preparation of genomic library of aureobasidin resistant strain having aureobasidin resistant gene *scaur2^R*

A genomic library was prepared from an aureobasidin resistant strain L22-8B by the same method as the one described in Example 2-c). E. coli containing this genomic library was cultured in an LB medium (50 ml) containing ampicillin and tetracycline at 37 °C overnight. Then plasmids were recovered and purified from the E. coli cells.

2-g) Expression and cloning of aureobasidin resistant gene *scaur2^R*

The above-mentioned plasmids originating in the genomic library of the L22-8B strain were transformed into S. cerevisiae SH3328 by the above-mentioned method of R.H. Schiestl. From the transformed strains, an aureobasidin resistant strain was isolated. Then a plasmid DNA containing the *scaur2^R* gene was recovered from this transformant by the above-mentioned method of I. Hagan et al. This plasmid was transformed into a yeast again and it was confirmed that the transformant had acquired aureobasidin resistance. This plasmid, which contained a DNA of 8.5 kb, was named pSCAR2. Fig. 3 shows the restriction enzyme map of the DNA fragment of 8.5 kb containing this aureobasidin resistant gene *scaur2^R*. E. coli HB101 having this plasmid pSCAR2 introduced therein was named and designated as Escherichia coli HB101 pSCAR2. This strain has been deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4484. By using BamHI, EcoRI, HindIII and PstI, DNA fragments of various sizes were prepared and cloned into the pWH5 vector. These plasmids were transformed into S. cerevisiae DKD5D in accordance with the above-mentioned method of R.H. Schiestl et al. Then it was examined whether these transformants had acquired aureobasidin resistance or not. As a result, none of the transformants of the DNA fragments was a resistant one. Thus it has been clarified that the DNA fragment of the full length is necessary for the expression of the aureobasidin resistance.

2-h) Isolation of aureobasidin sensitive gene *scaur2^S* corresponding to aureobasidin resistant gene *scaur2^R*

An E. coli stock containing the genomic library of Example 2-e) prepared from normal cells of S. cerevisiae DKD5D was spreaded on an LB agar medium containing ampicillin and tetracycline and incubated at 37 °C overnight. The colonies thus formed were transferred onto a nylon membrane (Hybond™-N) and the colony hybridization was performed. As a probe the DNA fragment of 8.5 kb obtained in the above Example 2-g) and labeled with [α -³²P] dCTP by using a random primer DNA labeling kit was used. As the results of screening of 2×10^4 colonies, several clones being hybridizable with the probe were obtained. Some of these clones contained a DNA fragment of 4.6 kb while others contained a DNA fragment of 3.9 kb. From the restriction enzyme maps of these DNA fragments, it was found out that these DNA fragments were each a part of the *scaur2^S* gene shown in Fig. 3. These DNA fragments were ligated together to thereby give a *scaur2^S* fragments shown in Fig. 3. The DNA fragment of 8.5 kb thus obtained was subcloned into pUC118 and then the DNA nucleotide sequence was determined (SEQ ID No. 9 in Sequence Listing). Based on the nucleotide sequence of SEQ ID No. 9 in Sequence Listing, the amino acid sequence represented by SEQ ID No. 10 in Sequence Listing was estimated.

Example 3: Gene disruption test on *spaur1^S* and *scaur1^S* genes

3-a) Gene disruption test on *spaur1^S* gene

In order to examine whether the aureobasidin sensitive gene *spaur1^S* is necessary in the cell growth by the gene disruption test, the plasmid pUARS2R prepared in Example 1-d) was first cleaved with BalI and EcoT22I. After eliminating a DNA fragment of 240 bp, the residual DNA fragment was blunted by using a

DNA blunting kit (manufactured by Takara Shuzo Co., Ltd.). Then this DNA was ligated with a DNA containing *ura4⁺* gene of 1.7 kb, which had been obtained by excising from a pUC8*ura4* plasmid [Mol. Gen. Genet., 215, 81 - 86 (1988)] by cleaving with *Hind*III and blunting, to thereby give a plasmid pUARS2RBT22::ura4-1 and another plasmid pUARS2RBT22::ura4-6 in which the *ura4* DNA had been inserted in the opposite direction. Both of these disrupted genes were excised from the vector pUC118 by cleaving with *Sac*I and *Hind*III and ARS2RBT22::ura4-1 and ARS2RBT22::ura4-6 (Fig. 4), which were *spaur1^S* DNA fragments containing *ura4⁺*, were purified. The purified DNA fragments were transformed into diploid cells *Schizo. pombe* C525 (*h⁹⁰ h⁹⁰*, *ura4-D18:ura4-D18*, *leu1 leu1*, *ade6-M210:ade6-M216*) by the above-mentioned method of Okazaki et al. and then a transformant was screened on an SD agar plate containing leucine. In the transformant thus obtained, one of a pair of *spaur1^S* genes on the chromosome had been replaced by the disrupted gene ARS2RBT22::ura4-1 or ARS2RBT22::ura4-6 introduced thereinto. These cells were allowed to undergo sporulation on a sporulation medium MEA and subjected to the tetrad analysis. As a result, it was found out that two of the four ascospores formed colonies but the residual two spores formed no colony. That is to say, the spores suffering from the replacement of the normal *spaur1^S* gene by the disrupted gene ARS2RBT22::ura4-1 were not propagated. It has been thus revealed that the *spaur1^S* gene is essentially required for the growth of the cells.

3-b) Gene disruption test on *scaur1^S* gene

The plasmid pSCAR1 prepared in Example 2-e) was partially digested with *Hind*III to thereby give a DNA fragment of 3.5 kb shown in Fig. 2. This DNA fragment was cloned into the *Hind*III site of pUC119 and the obtained product was named pSCAR3. The obtained pSCAR3 was cleaved with *Stu*I and *Eco*T22I. After eliminating a DNA fragment of 0.3 kb, the obtained DNA was ligated with a DNA fragment (1.1 kb) of *URA3* gene which had been obtained by cleaving a plasmid pYE*Ura3* (manufactured by Clontech Laboratories, Inc.) with *Hind*III and *Eco*RI and blunting. Thus a plasmid pUSCAR3.ST22::URA3⁺ and another plasmid pUSCAR3.ST22::URA3A, in which the *URA3* gene had been inserted in the opposite direction, were obtained. These disrupted gene were excised in the *Eco*RI site in the *scaur1^S* gene and the *Eco*RI site in the pUC119 vector by cleaving with *Eco*RI. The *scaur1^S* DNA fragments containing *URA3*, SCAR3.ST22::URA3⁺ and SCAR3.ST22::URA3A (Fig. 5), were purified. The purified DNA fragments were transformed into diploid cells of *S. cerevisiae* AOD1 (mating type *a/α*, genotype *ura3-52/ura3-52*, *leu2-3 112:leu2-3 112*, *trp1:TRP1*, *thr4:THR4*, *his4:HIS4*) by the above-mentioned method of R.H. Schiestl and transformants were screened on an SD agar plate containing leucine. The transformants thus obtained were allowed to undergo sporulation on a sporulation medium SP (1% of potassium acetate, 2% of agar) and subjected to the tetrad analysis. As a result, it was found out that two of the four ascospores underwent germination and formed colonies but the residual two spores did not undergo colony formation. That is to say, the spores suffering from the replacement of the *scaur1^S* gene by the disrupted gene were not propagated. It has been thus revealed that the *scaur1^S* gene is essentially required for the growth of the cells.

Example 4: Examination on the expression of aureobasidin sensitive gene *spaur1* by northern hybridization

From a normal strain or a resistant strain of *Schizo. pombe*, the whole RNAs were extracted and purified by the method of R. Jensen et al. [Proc. Natl. Acad. Sci. USA, 80, 3035 - 3039 (1983)]. Further, poly(A)⁺RNA was purified by using OligotexTM-dT30 (manufactured by Takara Shuzo Co., Ltd.). The purified poly(A)⁺RNA (2.5 μg) was separated by the electrophoresis on a 1.2% agarose gel containing formaldehyde and transferred onto a nylon membrane (HybondTM-N). After immobilizing, the hybridization was performed with the use of a *Hind*III-*Sac*I fragment (2 kb) of the *spaur1^R* gene labeled with [α -³²P]dCTP as a probe. As a result, both of the normal cells and the resistant cells showed a band of the same amount of about 2 kb. In both cases, this amount underwent no change in the logarithmic growth phase and the stationary phase (Fig. 10). Fig. 10 is an autoradiogram showing the results of the northern hybridization wherein mRNAs obtained from cells of a sensitive strain of *Schizo. pombe* in the logarithmic growth phase (lane 1), cells of a resistant strain in the logarithmic growth phase (lane 2), cells of the sensitive strain in the stationary phase (lane 3) and cells of the resistant strain in the stationary phase (lane 4) are electrophoresed on a 1.2% agarose gel containing formaldehyde.

Example 5: Determination of the activity of *scaur1^S* gene

5-a) Construction of plasmid YEpSCARW3 (Fig. 9) and YEpSCARW1

The plasmid pSCAR1 prepared in Example 2-e) was cleaved with *Hind*III and a fragment of 2 kb containing the whole ORF was excised. This fragment was inserted into the *Hind*III site of a expression-plasmid YEp52 having a promoter Gal10, the expression of which was induced by galactose in a medium. The plasmid having the *scaur1^S* gene which had been inserted in such a direction as to be normally transcribed by the promoter Gal10 was named YEpSCARW3. Fig. 9 shows the structure of this plasmid. Further, the plasmid having the *scaur1^S* gene inserted in the opposite direction was named YEpSCARW1.

5-b) Transformation by plasmids YEpSCARW3 and YEpSCARW1

By using 5 µg portions of the plasmids YEpSCARW3 and YEpSCARW1, the diploid *S. cerevisiae* cells with the disrupted *scaur1^S* gene prepared in Example 3-b) were transformed. Then transformants were screened on an SD agar plate. These transformants were allowed to undergo sporulation on an SP medium and then subjected to the tetrad analysis. When the expression of the *scaur1^S* gene was induced by using a YPGal medium (1% of yeast extract, 2% of polypeptone, 2% of galactose), the ascospores formed from the diploid cells transformed by YEpSCARW3 all underwent germination while two of the four ascospores formed from the diploid cells transformed by YEpSCARW1 underwent germination but not the remaining two. It is thus conceivable that the cells with the disrupted *scaur1^S* gene have reverted to the normal state by introducing YEpSCARW3 containing the *scaur1^S* gene into these cells. Accordingly, the use of these cells with the disrupted *scaur1^S* gene as a host makes it possible to determine the activity of normal *aur1*-analogous genes carried by other organisms.

Example 6: Confirmation and cloning of *aur1* and *aur2* genes (*caaur1*, *caaur2*) carried by *C. albicans*6-a) Detection of *aur1* gene by the PCR method

Poly(A)⁺RNA was extracted and purified from an aureobasidin sensitive strain *C. albicans* TIMM0136 by the same method as the one employed in Example 4. By using the poly(A)⁺RNA (5 µg) as a template, a double-stranded cDNA was synthesized on a cDNA synthesizing system Plus (manufactured by Amersham) with the use of an oligo(dT) primer. Mixed primers for PCR corresponding to amino acid sequence regions being common to the amino acid sequences of *S. cerevisiae* and *Schizo. pombe* were synthesized on a DNA synthesizer and purified. That is to say, a primer of SEQ ID No. 11 in Sequence Listing corresponding to the region of amino acids at the 184- to 192-positions of SEQ ID No. 4 in sequence Listing of *Schizo. pombe* (from the 184- to 192-positions of SEQ ID No. 8 in Sequence Listing of *S. cerevisiae*) and another primer of SEQ ID No. 12 in Sequence Listing corresponding to the region of amino acids from the 289- to 298-positions of *Schizo. pombe* (from the 289- to 298-positions of SEQ ID No. 8 in Sequence Listing of *S. cerevisiae*) were employed.

PCR was performed by using these primers and the above-mentioned cDNA as a template by repeating a cycle comprising treatment at 94°C for 30 seconds, one at 48°C for 1 minute and one at 72°C for 2 minutes 25 times. As a result, a DNA (about 350 bp) being almost the same as *S. cerevisiae* and *Schizo. pombe* in length was amplified (Fig. 6). Fig. 6 shows a pattern obtained by carrying out PCR with the use of cDNA of *C. albicans* (lane 1), cDNA of *S. cerevisiae* (lane 2) and cDNA of *Schizo. pombe* (lane 3) as a template, electrophoresing each PCR product on an agarose gel and staining with ethidium bromide.

6-b) Cloning of *aur1* gene (*caaur1*) of *C. albicans*

(i) Genomic DNA was extracted and purified from a strain *C. albicans* TIMM0136 by the same method as the one described in Example 1-c). After partially digesting with *Hind*III, the DNA fragment was ligated with a TrapexII9 vector which had been completely digested with *Hind*III and transformed into *E. coli* HB101. Thus a genomic library of *C. albicans* was prepared. From this library, a DNA fragment of 4.5 kb containing the *aur1* gene of *C. albicans* was cloned by using the DNA fragment of *C. albicans* obtained by the PCR described in Example 6-a), which had been labeled with [α -³²P]dCTP by using a random primer DNA labeling kit (manufactured by Takara Shuzo Co., Ltd.), as a probe. This DNA fragment had a restriction enzyme map shown in Fig. 7 and the DNA nucleotide sequence thereof is represented by SEQ ID No. 13 in Sequence Listing. Based on this nucleotide sequence, it was estimated that the *caaur1*

gene coded for a protein having the amino acid sequence represented by SEQ ID No. 14 in Sequence Listing. When compared with the *scaur1^S* protein, a homology of as high as 53% was observed. A TraplexII9 vector having this *caaur1* gene integrated therein was named pCAAR1, while *E. coli* HB101 transformed by this plasmid was named and designated as *Escherichia coli* HB101/pCAAR1. This strain has been deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4482.

Next, pCAAR1 was treated with *Hind*III to thereby give *caaur1* of 4.5 kb. Further, it was integrated into pTV118 which had been completely digested with *Hind*III to thereby prepare a plasmid for expressing *caaur1*. This plasmid was named pTCAAR1.

(ii) Genomic DNA was extracted and purified from a strain *C. albicans* TIMM1768 [The journal of Antibiotics, 46, 1414-1420(1993)] by the same method as the one described in example 1-c). After partially digesting with *Hind* III, the DNA fragment was ligated with a pUC118 vector which had been completely digested with *Hind* III and transformed into *E. coli* HB101. Thus a genomic library of *C. albicans* TIMM1768 was prepared. From this library, a DNA fragment of 4.5 kb containing the *aur1* gene of *C. albicans* TIMM1768 was cloned by the colony hybridization with the same probe as that described in Example 6-b)-(i). This DNA fragment had the same restriction enzyme map as that shown in Fig. 7. Next, a part of the DNA sequence containing a ORF in this DNA fragment was determined. The DNA nucleotide sequence thereof is represented by SEQ ID No. 21 in Sequence Listing. Based on this nucleotide sequence, it was estimated that this gene coded for a protein having the amino acid sequence represented by SEQ ID No. 22 in Sequence Listing. When the amino acid sequence of the *caaur1* protein *C. albicans* TIMM1768 was compared with that of the *caaur1* protein of *C. albicans* TIMM0136, the amino acid sequences of the 1- to 381-positions and the 383- to 423-positions and the 425- to 471-positions of *caaur1* protein (SEQ ID No. 14 in Sequence Listing) in *C. albicans* TIMM0136 were identical with the amino acid sequences of the 2- to 382-positions and the 384- to 424-positions and the 426- to 472-positions, respectively, of *caaur1* protein (SEQ ID No. 22 in Sequence Listing) in *C. albicans* TIMM1768.

However, serines at the 382- and 424-positions of SEQ ID No. 14 in Sequence Listing were replaced with prolines at the 383- and 425-positions of SEQ ID No. 22 in Sequence Listing.

6-c) Cloning of *aur2* gene (*caaur2*) of *C. albicans*

Genomic DNA of a strain *C. albicans* TIMM0136 was digested with *Bam*HI and ligated with a pTV118 vector which had been completely digested with *Bam*HI. Then it was transformed into *E. coli* HB101 to thereby prepare a genomic library of *C. albicans*. On the other hand, the DNA fragment containing the *scaur2^S* gene obtained in Example 2-h) was cleaved with *Hind*III and *Pst*I to thereby give a DNA fragment of 1.2 kb. This DNA fragment was labeled with [α -³²P]dCTP by using a random primer DNA labeling kit. By using this labeled DNA fragment as a probe, the above-mentioned *C. albicans* genomic library was screened by the colony hybridization. Thus a plasmid containing a DNA fragment of 8.3 kb was obtained. A part of the DNA sequence upstream of the *Bam*HI site of this DNA fragment was determined (SEQ ID No. 15 in Sequence Listing). Based on this sequence, an amino acid sequence represented by SEQ ID No. 16 in Sequence Listing was estimated. It corresponded to the amino acid sequence of the 1230- to 1309-positions of the amino acid sequence of the *scaur2* gene (SEQ ID No. 10), having a homology of as high as 77%. Since this DNA fragment lacked a part of the C-end, the genomic library prepared in Example 6-b) was further screened by using this DNA fragment as a probe. Thus a DNA fragment of 6.5 kb having the C-terminal part was obtained. Fig. 8 shows the restriction enzyme map of the DNA region containing the *caaur2* gene thus clarified.

A pTV118 vector having the above-mentioned *caaur2* gene of 8.3 kb integrated therein was named pCAAR2N, while *E. coli* HB101 transformed by this plasmid was named and designated as *Escherichia coli* HB101/pCAAR2N. This strain has been deposited at National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology in accordance with the Budapest Treaty under the accession number FERM BP-4481.

Example 7: Preparation of antibody against protein coded for by *scaur1^S* gene and staining of *S. cerevisiae* cells and detection of said protein by using this antibody.

7-a) Preparation of antibody

SCAR1-1 (SEQ ID No. 19 in Sequence Listing) comprising a peptide corresponding to the amino acids at the residue 103 to 113 in the amino acid sequence of SEQ ID No. 8 in Sequence Listing having cysteine added to the N-end thereof and SCAR1-2 (SEQ ID No. 20 in Sequence Listing) comprising a peptide corresponding to the amino acids at the residue 331 to 348 in the amino acid sequence of SEQ ID No. 8 having cysteine added to the N-end thereof were synthesized by the Fmoc solid phase synthesis method and purified by reverse phase HPLC. Thus 10 mg portions of these peptides were obtained. To the N-terminal cysteine of each of these synthetic peptides, KLH was bound as a carrier protein. By using this binding product as an antigen, a rabbit was immunized and an antiserum was obtained. This antiserum was further purified on an affinity column prepared by binding the synthetic peptide employed as the antigen to an agarose gel. This a polyclonal antibody being specific for the synthetic peptide was prepared.

7-b) Staining of *S. cerevisiae* cells with antibody

A strain *S. cerevisiae* ATCC 9763 was cultured in a YNBG medium [0.67% of yeast nitrogen base (manufactured by Difco), 2% of glucose] to thereby give a suspension of a concentration of 3×10^7 cells/ml. To 1 ml of this cell suspension were added 0.11 ml of a 1 M phosphate buffer (pH 6.5) and 0.17 ml of 37% formaldehyde. After slowly stirring at room temperature for 1 hour, the cells were harvested by centrifugation and then suspended in 20 ml of an SS buffer (1 M of sorbitol, 0.2 % of β -mercaptoethanol, 0.1 M phosphate buffer, pH 7.5) containing 20 μ g/ml of Zymolyase 20T. After treating at 30 °C for 1 hour, the cells were harvested, washed with the SS buffer, suspended in 1 ml of the SS buffer containing 0.1% of Triton X-100 and then allowed to stand for 10 minutes. This cell suspension was placed on a slide glass which had been coated with poly(L-lysine) and allowed to stand for 10 minutes. Next, a PBS solution containing 1% of albumin (BSA) was dropped thereinto. After allowing to stand at room temperature for 15 minutes, the excessive liquid was removed and then a PBS solution containing BSA containing 0.02 mg/ml of the antiSCAR1-1 antibody was dropped thereinto. After allowing to stand at room temperature for 60 minutes and washing with PBS containing BSA three times, antirabbit IgG antibody labeled with FITC (antibody concentration 0.02 mg/ml) was layered over and allowed to stand at room temperature for 1 hour. After washing with a PBS solution containing BSA, a small amount of a mounting solution, which was a solution prepared by dissolving 0.1 g of p-phenylenediamine in 10 ml of CBS (150 mM of NaCl, 50 mM of CHES, pH 9.5), adjusting the pH value to 9.0 with 10 N NaOH and further adding 90 ml of glycerol, was layered over. Then a cover glass was placed thereon to thereby give a specimen. This specimen was observed under a fluorescence microscope to thereby examine the intracellular distribution of the *scaur1* protein. As a result, it was found out that this protein was distributed all over the cells.

7-c) Detection of protein coded for by *scaur1* gene by using antibody

The plasmid YEpSCARW3 prepared in Example 5-a) was introduced into a normal haploid *S. cerevisiae* SH3328 to thereby give a transformant. This transformant was cultured in a YPGal medium or a YPD medium and the cells were harvested by centrifugation. The cells thus obtained were suspended in a buffer (1% of Triton X-100, 1% of SDS, 20 mM of Tris-HCl, pH 7.9, 10 mM of EDTA, 1 mM of DTT, 1 mM of PMSF). Further, glass beads were added thereto to disrupt the cells by vigorous vortex. Then an SDS loading solution was added thereto and the protein was denatured by treating at 95 °C for 5 minutes. After centrifuging, a part of the obtained supernatant was subjected to SDS-PAGE and the protein thus separated was transferred onto an Immobilon membrane (manufactured by MILLIPORE). This Immobilon membrane was treated with Block Ace (manufactured by Dainippon Pharmaceutical Co., Ltd.). Then the antiSCAR1-2 antibody prepared in 7-a) was reacted therewith as a primary antibody. After washing, antirabbit IgG antibody labeled with peroxidase was reacted therewith as a secondary antibody and the mixture was thoroughly washed. Next, it was color-developed with diaminobenzidine and a band of the *scaur1* protein was detected. Fig. 11 shows the results.

Fig. 11 shows the results of the detection of the protein prepared from the cells incubated in the YPD medium (lane 1) and the protein prepared from the cells incubated in the YPGal medium (lane 2), each subjected to SDS-PAGE, by using the antiSCAR1-2 antibody. The cells incubated in the YPGal medium, of which *scaur1* gene had been induced, showed a specific band.

[Effects of the Invention]

According to the present invention, a novel protein regulating aureobasidin sensitivity and a gene coding for the protein, i.e., a gene regulating aureobasidin sensitivity are provided. These substances are useful in the diagnosis and treatment for diseases caused by organisms having the above-mentioned gene, such as mycoses. The present invention further provides an antisense DNA and an antisense RNA of this gene, a nucleic acid probe being hybridizable with the gene, a process for detecting the gene by using this nucleic acid probe, a process for producing a protein regulating aureobasidin sensitivity by using a transformant having the gene introduced therein, an antibody for the protein and a process for detecting the protein by using this antibody. They are also useful in the diagnosis and treatment of diseases including mycoses.

15

20

25

30

35

40

45

50

55

Sequence Listing

SEQ ID NO : 1

SEQUENCE LENGTH : 2385

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

AAGCTTTTTT GCCTCTGCAA AAGTTCCTTT CTCGAATTGG TTTTTTGAGG AAAAGCAAGT 60
TAATAAACTA ATTATATTAT ATATAATTAG CAATTTTATA AAAAAAATAA AAAAATAGCC 120
CTGATTGCTG GCAACTGTGA GCTGAACATT GGTAAATCGG TCCATCTTTT TTAAATATT 180
TTACATCGCT ACTTTTAAAGT GCTTGACACT TGCATTTAAT AGCTACTTTC TTCCCTTCAT 240
AAAAATTCCT TTTTTTCTT TTAGTTTTCC GGTAAATTCC TTACGAAATT TTTTTCGTAC 300
GCTTCCCTTT TTTACTCTGA TAATTCTTTG AAGCAATGTC TGCTCTTTCG ACCTTAAAAA 360
AGCGCCTTGC TGGGTGTAAC CGAGCATCCC AATACAAGTT GGAAACAAGC TTAAACCCTA 420
TGCCTACATT TCGTTTGCTA CGCAATACGA AATGGTCATG GACACATTTG CAATATGTGT 480
TTCTAGCAGG TAATTTGATT TTTGCTTGTA TTGTCATTGA ATCTCCTGGA TTCTGGGGGA 540
AATTTGGCAT TGCCTGTCTT TTGGCCATTG CGTTGACCGT TCCTTTAACA CGCCAAATTT 600
TTTTTCCTGC CATTGTTATC ATCACCTGGG CAATTTTATT TTACTCTTGT AGGTTTATTC 660
CAGAACGCTG GCGTCCACCC ATATGGGTTT GTGTTTTACC CACACTTGAA AATATTCTTT 720
ATGGCTCTAA TCTTTCTAGT CTTCTCTCGA AAACCACGCA TAGCATCCTT GATATTTTGG 780
CCTGGGTTCC ATATGGAGTC ATGCATTATT CGGCTCCTTT TATCATTTCA TTTATTCTTT 840
TCATCTTTGC ACCTCCTGGA ACTCTCCAG TTTGGGCTCG AACTTTTGGT TATATGAATT 900
TATTTGGTGT TCTTATCCAA ATGGCTTTCC CCTGTTCTCC TCCTTGGTAT GAAAATATGT 960
ATGGTTTAGA ACCTGCCACG TATGCAGTAC GTGGCTCTCC TGGTGGATTG GCCCGTATTG 1020
ATGCTCTCTT CGGCACTAGC ATTTACACTG ATTGTTTTTC TAACTCTCCG GTTGTTTTTG 1080
GTGCCTTTCC ATCTCTTAC GCTGGATGGG CCATGCTGGA AGCACTTTTC CTTTCGCATG 1140
TGTTTCCTCG ATACCGCTTC TGCTTTTATG GATATGTTCT ATGGCTTTGC TGGTGTACTA 1200
TGTACCTTAC CCACCACTAC TTTGTAGATT TGGTCGGCGG TATGTGTTTA GCTATTATAT 1260

GCTTCGTTTT TGCTCAAAAG CTACGCCTCC CACAGTTGCA AACTGGTAAA ATCCTTCGTT 1320
 GGGAAATACGA GTTTGTTATC CACGGTCATG GTCCTTCCGA AAAAACCAGC AACTCCTTGG 1380
 5 CTCGTACCGG CAGCCCATAC TTA CTCTGGAA GGGATTCTTT TACTCAAAAC CCTAATGCAG 1440
 TAGCCTTCAT GAGTGGTCTT AACAATATGG AACTTGCTAA CACCGATCAT GAATGGTCCG 1500
 TGGGTTCATC ATCACCTGAG CCGTTACCTA GTCCTGCTGC TGATTTGATT GATCGTCCTG 1560
 10 CCAGTACCAC TTCCTCCATC TTTGATGCAA GTCATCTTCC TTAAATCAAC GTGCTTTAAG 1620
 AATATATTTT CAAAAGCTAC ATGATACATT GACTAGAATC GGTTTGATTG ATAGTGGTAT 1680
 TGAATGATG TTGTTTATTG TGT TTTTAA CTGTTAATCT GACATCCATT GAGTCATTCT 1740
 15 TTACAATTTG TAAAATTAAT TTGTATCACT AATTTTGAAG GAAGCTATTT TGGTATTAAT 1800
 ACCGCTTTTG GTCTCCACTT CCTTTTCGAA ACTCTTAACA GCGATTAGGC CGGGTATCTT 1860
 CCAGTGTGAT GTATAGGTAT TTGTCGTTTT TTTATCATTT CCGTTAATAA AGAACTCTTT 1920
 20 TATCCAGCTT CTTACACTGT CAACTGTTGT GAAAGGAACA CATTTAGAAT TTCATTTTCC 1980
 TTATTTGTTG TGATTTAAAT CGTTTGACAT AATTTTAAAT TTGGTTTGAA ATGTGTGTGA 2040
 GAAGGCTTGT TTTATTCATT TAGTTTATTG CTGTTTGCA CGAAAATCCA GAACGGAGCA 2100
 25 TTAATGTAAT CCTTTTTTAT TCTGTAAAGC GTTTTATAC AAATGTTGGT TATACGTTTC 2160
 TAAAATAAGA ATATTGTTAT AATAATATAG TTTTCTAT CATTTGTTAC ACACACTAAA 2220
 GAGACATTAA GGATAAGCAA ATGTGTTAAA ATGATAATAT ATTTTGAAA CATTTATAAA 2280
 30 GAAATTAAGC AGCTTTGACT AACTACATTT TTGTTTTTTT CCTAAGCAAA ACTGTATAGT 2340
 TATACACGCG AGCTGTATTC ACTTCCATTG TAGTGACTTG AGCTC 2385

35 SEQ ID NO : 2

SEQUENCE LENGTH : 422

40 SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

45 MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Met Ser Ala Leu Ser Thr Leu Lys Lys Arg Leu Ala Ala Cys Asn

50 1 5 10 15

55

EP 0 644 262 A2

	Arg Ala Ser Gln Tyr Lys Leu Glu Thr Ser Leu Asn Pro Met Pro	
	20	25 30
5	Thr Phe Arg Leu Leu Arg Asn Thr Lys Trp Ser Trp Thr His Leu	
	35	40 45
10	Gln Tyr Val Phe Leu Ala Gly Asn Leu Ile Phe Ala Cys Ile Val	
	50	55 60
	Ile Glu Ser Pro Gly Phe Trp Gly Lys Phe Gly Ile Ala Cys Leu	
15	65	70 75
	Leu Ala Ile Ala Leu Thr Val Pro Leu Thr Arg Gln Ile Phe Phe	
	80	85 90
20	Pro Ala Ile Val Ile Ile Thr Trp Ala Ile Leu Phe Tyr Ser Cys	
	95	100 105
	Arg Phe Ile Pro Glu Arg Trp Arg Pro Pro Ile Trp Val Arg Val	
25	110	115 120
	Leu Pro Thr Leu Glu Asn Ile Leu Tyr Gly Ser Asn Leu Ser Ser	
	125	130 135
30	Leu Leu Ser Lys Thr Thr His Ser Ile Leu Asp Ile Leu Ala Trp	
	140	145 150
	Val Pro Tyr Gly Val Met His Tyr Ser Ala Pro Phe Ile Ile Ser	
35	155	160 165
	Phe Ile Leu Phe Ile Phe Ala Pro Pro Gly Thr Leu Pro Val Trp	
	170	175 180
40	Ala Arg Thr Phe Gly Tyr Met Asn Leu Phe Gly Val Leu Ile Gln	
	185	190 195
	Met Ala Phe Pro Cys Ser Pro Pro Trp Tyr Glu Asn Met Tyr Gly	
45	200	205 210
	Leu Glu Pro Ala Thr Tyr Ala Val Arg Gly Ser Pro Gly Gly Leu	
	215	220 225
50	Ala Arg Ile Asp Ala Leu Phe Gly Thr Ser Ile Tyr Thr Asp Cys	

55

EP 0 644 262 A2

	230	235	240
	Phe Ser Asn Ser Pro Val Val Phe Gly Ala Phe Pro Ser Leu His		
5	245	250	255
	Ala Gly Trp Ala Met Leu Glu Ala Leu Phe Leu Ser His Val Phe		
10	260	265	270
	Pro Arg Tyr Arg Phe Cys Phe Tyr Gly Tyr Val Leu Trp Leu Cys		
	275	280	285
15	Trp Cys Thr Met Tyr Leu Thr His His Tyr Phe Val Asp Leu Val		
	290	295	300
	Gly Gly Met Cys Leu Ala Ile Ile Cys Phe Val Phe Ala Gln Lys		
20	305	310	315
	Leu Arg Leu Pro Gln Leu Gln Thr Gly Lys Ile Leu Arg Trp Glu		
	320	325	330
25	Tyr Glu Phe Val Ile His Gly His Gly Leu Ser Glu Lys Thr Ser		
	335	340	345
	Asn Ser Leu Ala Arg Thr Gly Ser Pro Tyr Leu Leu Gly Arg Asp		
30	350	355	360
	Ser Phe Thr Gln Asn Pro Asn Ala Val Ala Phe Met Ser Gly Leu		
	365	370	375
35	Asn Asn Met Glu Leu Ala Asn Thr Asp His Glu Trp Ser Val Gly		
	380	385	390
	Ser Ser Ser Pro Glu Pro Leu Pro Ser Pro Ala Ala Asp Leu Ile		
40	395	400	405
	Asp Arg Pro Ala Ser Thr Thr Ser Ser Ile Phe Asp Ala Ser His		
45	410	415	420
	Leu Pro		

SEQ ID NO : 3

SEQUENCE LENGTH : 2385

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

AAGCTTTTT GCCTCTGCAA AAGTTCCTTT CTCGAATTGG TTTTTTGAGG AAAAGCAAGT 60
 TAATAAACTA ATTATATTAT ATATAATTAG CAATTTTATA AAAAAAATAA AAAAATAGCC 120
 CTGATTGCTG GCAACTGTGA GCTGAACATT GGTTAATCGG TCCATCTTTT TTAAATATT 180
 TTACATCGCT ACTTTTAAGT GCTTGACACT TGCATTTAAT AGCTACTTTC TTTCCTTCAT 240
 AAAAAATCCT TTTTTTTCCT TTAGTTTTCC GGTTAATTCC TTACGAAATT TTTTTCGTAC 300
 GCTTCCCTTT TTTACTCTGA TAATTCCTTG AAGCAATGTC TGCTCTTTCG ACCTTAAAAA 360
 AGCGCCTTGC TCGGTGTAAC CGAGCATCCC AATACAAGTT GGAAACAAGC TTAAACCCTA 420
 TGCCTACATT TCGTTTGCTA CGCAATACGA AATGGTCATG GACACATTTG CAATATGTGT 480
 TTCTAGCAGG TAATTTGATT TTTGCTTGTA TTGTCATTGA ATCTCCTGGA TTCTGGGGGA 540
 AATTTGGCAT TGCCTGTCTT TTGGCCATTG CGTTGACCGT TCCTTTAACA CGCCAAATTT 600
 TTTTTCCTGC CATTGTTATC ATCACCTGGG CAATTTTATT TTA CTCTGTG AGGTTTATTC 660
 CAGAACGCTG GCGTCCACCC ATATGGGTTT GTGTTTTACC CACACTTGAA AATATTCTTT 720
 ATGGCTCTAA TCTTCTAGT CTTCTCTCGA AAACCACGCA TAGCATCCTT GATATTTTGG 780
 CCTGGGTTCC ATATGGAGTC ATGCATTATT CGGCTCCTTT TATCATTTC A TTTATTCTTT 840
 TCATCTTTGC ACCTCCTGGA ACTCTTCCAG TTTGGGCTCG AACTTTTGGT TATATGAATT 900
 TATTTGGTGT TCTTATCCAA ATGGCTTTCC CCTGTTCTCC TCCTTGGTAT GAAAAATATGT 960
 ATGGTTTAGA ACCTGCCACG TATGCAGTAC GTGGCTCTCC TGGTGGATTG GCCCGTATTG 1020
 ATGCTCTCTT CGGCACTAGC ATTTACACTG ATGGTTTTTC TAACTCTCCG GTTGTTTTTG 1080
 GTGCCTTTCC ATCTCTTAC GCTGGATGGG CCATGCTGGA AGCACTTTTC CTTTCGCATG 1140
 TGTTCCTCG ATACCGCTTC TGCTTTTATG GATATGTTCT ATGGCTTTCG TGGTGTACTA 1200
 TGTACCTTAC CCACCACTAC TTTGTAGATT TGGTCGGCGG TATGTGTTTA GCTATTATAT 1260
 GCTTCGTTTT TGCTCAAAAG CTACGCCTCC CACAGTTGCA AACTGGTAAA ATCCTTCGTT 1320
 GGGAATACGA GTTTGTTATC CACGGTCATG GTCTTTCCGA AAAAACCAGC AACTCCTTGG 1380
 CTCGTACCGG CAGCCCATAC TTA CTTGAA GGGATTCTTT TACTCAAAAC CCTAATGCAG 1440

TAGCCTTCAT GAGTGGTCTT AACAAATATGG AACTTGCTAA CACCGATCAT GAATGGTCCG 1500
 TGGGTTTCATC ATCACCTGAG CCGTTACCTA GTCCTGCTGC TGATTGATT GATCGTCCTG 1560
 5 CCAGTACCAC TTCCTCCATC TTTGATGCAA GTCATCTTCC TTAAATCAAC GTGCTTTAAG 1620
 AATATATTTT CAAAAGCTAC ATGATACATT GACTAGAATC GGTGTTGATTC ATAGTGGTAT 1680
 10 TGAATGATG TTGTTTCATTG TGTTTTTTAA CTGTTAATCT GACATCCATT GAGTCATTCT 1740
 TTACAATTTG TAAAATTAAT TTGTATCACT AATTTTGAAG GAAGCTATTT TGGTATTAAT 1800
 ACCGCTTTTG GTCTCCACTT CCTTTTCGAA ACTCTTAACA GCGATTAGGC CGGGTATCTT 1860
 15 CCAGTGTGAT GTATAGGTAT TTGTCGTTTT TTTATCATTT CCGTTAATAA AGAACTCTTT 1920
 TATCCAGCTT CTTACACTGT CAACTGTTGT GAAAGGAACA CATTTAGAAT TTCATTTTCC 1980
 TTATTTGTTG TGATTTAAAT CGTTTGACAT AATTTTAAAT TTGGTTTGAA ATGTGTGTGA 2040
 20 GAAGGCTTGT TTTATTCATT TAGTTTATTG CTGTTTGCA CGAAAATCCA GAACGGAGCA 2100
 TTAATGTAAT CCTTTTTTAT TCTGTAAAGC GTTTTTATAC AAATGTTGGT TATACGTTTC 2160
 TAAAATAAGA ATATTGTTAT AATAATATAG TTTTTTCTAT CATTTGTTAC ACACACTAAA 2220
 25 GAGACATTAA GGATAAGCAA ATGTGTTAAA ATGATAATAT ATTTTGAAA CATTTATAAA 2280
 GAAATTAAGC AGCTTTGACT AACTACATTT TTGTTTTTTT CCTAAGCAAA ACTGTATAGT 2340
 TATACACGGC AGCTGTATTC ACTTCCATTG TAGTGACTTG AGCTC 2385

SEQ ID NO : 4

SEQUENCE LENGTH : 422

35 SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

40 MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Met Ser Ala Leu Ser Thr Leu Lys Lys Arg Leu Ala Ala Cys Asn

45 1 5 10 15

Arg Ala Ser Gln Tyr Lys Leu Glu Thr Ser Leu Asn Pro Met Pro

20 25 30

50 Thr Phe Arg Leu Leu Arg Asn Thr Lys Trp Ser Trp Thr His Leu

55

EP 0 644 262 A2

	35	40	45
	Gln Tyr Val Phe Leu Ala Gly Asn Leu Ile Phe Ala Cys Ile Val		
5	50	55	60
	Ile Glu Ser Pro Gly Phe Trp Gly Lys Phe Gly Ile Ala Cys Leu		
	65	70	75
10	Leu Ala Ile Ala Leu Thr Val Pro Leu Thr Arg Gln Ile Phe Phe		
	80	85	90
	Pro Ala Ile Val Ile Ile Thr Trp Ala Ile Leu Phe Tyr Ser Cys		
15	95	100	105
	Arg Phe Ile Pro Glu Arg Trp Arg Pro Pro Ile Trp Val Arg Val		
	110	115	120
20	Leu Pro Thr Leu Glu Asn Ile Leu Tyr Gly Ser Asn Leu Ser Ser		
	125	130	135
	Leu Leu Ser Lys Thr Thr His Ser Ile Leu Asp Ile Leu Ala Trp		
	140	145	150
	Val Pro Tyr Gly Val Met His Tyr Ser Ala Pro Phe Ile Ile Ser		
30	155	160	165
	Phe Ile Leu Phe Ile Phe Ala Pro Pro Gly Thr Leu Pro Val Trp		
	170	175	180
35	Ala Arg Thr Phe Gly Tyr Met Asn Leu Phe Gly Val Leu Ile Gln		
	185	190	195
	Met Ala Phe Pro Cys Ser Pro Pro Trp Tyr Glu Asn Met Tyr Gly		
40	200	205	210
	Leu Glu Pro Ala Thr Tyr Ala Val Arg Gly Ser Pro Gly Gly Leu		
	215	220	225
45	Ala Arg Ile Asp Ala Leu Phe Gly Thr Ser Ile Tyr Thr Asp Gly		
	230	235	240
	Phe Ser Asn Ser Pro Val Val Phe Gly Ala Phe Pro Ser Leu His		
50	245	250	255
55			

	Ala Gly Trp Ala Met Leu Glu Ala Leu Phe Leu Ser His Val Phe		
		260	265 270
5	Pro Arg Tyr Arg Phe Cys Phe Tyr Gly Tyr Val Leu Trp Leu Cys		
		275	280 285
10	Trp Cys Thr Met Tyr Leu Thr His His Tyr Phe Val Asp Leu Val		
		290	295 300
	Gly Gly Met Cys Leu Ala Ile Ile Cys Phe Val Phe Ala Gln Lys		
15		305	310 315
	Leu Arg Leu Pro Gln Leu Gln Thr Gly Lys Ile Leu Arg Trp Glu		
		320	325 330
20	Tyr Glu Phe Val Ile His Gly His Gly Leu Ser Glu Lys Thr Ser		
		335	340 345
	Asn Ser Leu Ala Arg Thr Gly Ser Pro Tyr Leu Leu Gly Arg Asp		
25		350	355 360
	Ser Phe Thr Gln Asn Pro Asn Ala Val Ala Phe Met Ser Gly Leu		
		365	370 375
30	Asn Asn Met Glu Leu Ala Asn Thr Asp His Glu Trp Ser Val Gly		
		380	385 390
	Ser Ser Ser Pro Glu Pro Leu Pro Ser Pro Ala Ala Asp Leu Ile		
35		395	400 405
	Asp Arg Pro Ala Ser Thr Thr Ser Ser Ile Phe Asp Ala Ser His		
		410	415 420
40	Leu Pro		

SEQ ID NO : 5

SEQUENCE LENGTH : 2340

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

5 TTTCTTTCTG TCAAAGAATA ATAAAGTGCC CATCAGTGTT CATATTTGTT ACAAAGTGGT 60
 TTTCTGATTT GGTACTACTG CAGAGGCGTA TTTTGTGCTT CAGTTACCAT AGCGTAAGAA 120
 10 CACTAGCGAC TTTTGTTCGT GAACCAACAG AGTAGGATTT CTACTGCTAC ATCTCTTAGG 180
 TAGTTGGTTA GTCCGATCGC TCACTTTGGG TTGTTGTAA GTACTTCATA AGTTTATCCT 240
 TTTCTTTTTT CACACTGAGC TACTTTGGGT ATAGCTTTTG GCCCAAGGAT CTTTGAATTT 300
 15 TCTCCAAAAG TACTTTATTT TATATCCTAC AGGTTGCGGT TTTCATATTT TAAAAAGCTT 360
 TTTAATCATT CCTTTGCGTA TGGCAAACCC TTTTTCGAGA TGGTTTCTAT CAGAGAGACC 420
 TCCAAACTGC CATGTAGCCG ATTTAGAAAC AAGTTTAGAT CCCCATCAA CGTTGTTGAA 480
 20 GGTGCAAAAA TACAAACCCG CTTTAAGCGA CTGGGTGCAT TACATCTTCT TGGGATCCAT 540
 CATGCTGTTT GTGTTTATTA CTAATCCCGC ACCTTGGATC TTCAAGATCC TTTTTTATTG 600
 TTTCTTGGGC ACTTTATTCA TCATTCCAGC TACGTCACAG TTTTCTTCA ATGCCTTGCC 660
 25 CATCCTAACA TGGGTGGCGC TGTATTTAC TTCATCGTAC TTTCCAGATG ACCGCAGGCC 720
 TCCTATTACT GTCAAAGTGT TACCAGCGGT GGAAACAATT TTATACGGCG ACAATTTAAG 780
 TGATATTCTT GCAACATCGA CGAATTCCTT TTTGGACATT TTAGCATGGT TACCGTACGG 840
 30 ACTATTTTAT TATGGGGCCC CATTTGTCGT TGCTGCCATC TTATTCGTAT TTGGTCCACC 900
 AACTGTTTTG CAAGGTTATG CTTTTGCATT TGTTATATG AACCTGTTTG GTGTTATCAT 960
 GCAAAATGTC TTTCCAGCCG CTCCCCCATG GTATAAAATT CTCTATGGAT TGCAATCAGC 1020
 35 CAACTATGAT ATGCATGGCT CGCCTGGTGG ATTAGCTAGA ATTGATAAGC TACTCGGTAT 1080
 TAATATGTAT ACTACAGCTT TTTCAAATTC CTCGTCATT TTCGGTGCTT TTCCTTCACT 1140
 GCATTCCGGG TGTGCTACTA TGGAAGCCCT GTTTTCTGT TATTGTTTTT CAAAATTGAA 1200
 40 GCCCTTGTTT ATTGCTTATG TTTGCTGGTT ATGGTGGTCA ACTATGTATC TGACACACCA 1260
 TTATTTTGTA GACCTTATGG CAGGTTCTGT GCTGTATAC GTTATTTTCC AGTACACAAA 1320
 GTACACACAT TTACCAATTG TAGATACATC TCTTTTTTGC AGATGGTCAT ACACTTCAAT 1380
 45 TGAGAAATAC GATATATCAA AGAGTGATCC ATTGGCTGCA GATTCAAACG ATATCGAAAG 1440
 TGTCCCTTTG TCCAACTTGG AACTTGACTT TGATCTTAAT ATGACTGATG AACCCAGTGT 1500
 50 AAGCCCTTCG TTATTTGATG GATCTACTTC TGTTCCTCGT TCGTCCGCCA CGTCTATAAC 1560
 GTCACTAGGT GTAAAGAGGG CTTAATGAGT ATTTTATCTG CAATTACGGA TACGGTTGGT 1620

55

CTTATGTAGA TACATATAAA TATATATCTT TTTCTTTCTT TTTCTTAGTC AGGATTGTCTG 1680
 TTTAGCATAA TATACATGTA GTTTATTTAA TCACATACCA CTGATTATCT TTAGAATTTT 1740
 ATAAATTTTT GAAATAAATG GGTGGCTTTT AATGGTGTCT ATGTTAAGTG AGGCTTTTATG 1800
 AATGCTCTTC CTGCTTTGTT TATTATATGT GTATGAAAGA TATGTATGTA TTTACATGTG 1860
 TTTGTAGCGT CCCCAGTCAA AACCTGTGCG CTATACCTAA ATGGATTGAT AATCTTCATT 1920
 CACTAATTCT AAAATAGACT TCTTCCCCAA AGAACGGTGT AACGATGAGG CTCTATCCAG 1980
 CTGCTTATCT AAATCAACTT TAACGATGGA TGATCTTATG ACACGGGGAT CTTTCTTTAA 2040
 AGTTCTTAGA ATTTTCAGACT GTACCGCAGC TGATGAATCA AACAGCATTA AAAAGTGATA 2100
 TGCTCGAAAA TGTTTTCTT GGTCTTTCTT CATTATTTTA GGAAGATACC TTATGCCCAT 2160
 GGGTACAATG TCCCTCACCA CACCTCTGTT TTGAATAATC AGTTTCCCGA TTGTGGAAGA 2220
 CAATTCTTTT GCTTCCAAC TGGGCGCATT GGAGTTGGTT ATGCGAACAA GTCCGATCAG 2280
 CTCATAAAGC ATCTTAGTGA AAAGGGTGGT TTTGCGTTAT TCTTTCCTCT GTTGAAGCTT 2340

SEQ ID NO : 6

SEQUENCE LENGTH : 401

SEQUENCE TYPE : amino acid

STRANDED : single

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Met Ala Asn Pro Phe Ser Arg Trp Phe Leu Ser Glu Arg Pro Pro

1 5 10 15

Asn Cys His Val Ala Asp Leu Glu Thr Ser Leu Asp Pro His Gln

20 25 30

Thr Leu Leu Lys Val Gln Lys Tyr Lys Pro Ala Leu Ser Asp Trp

35 40 45

Val His Tyr Ile Phe Leu Gly Ser Ile Met Leu Phe Val Phe Ile

50 55 60

Thr Asn Pro Ala Pro Trp Ile Phe Lys Ile Leu Phe Tyr Cys Phe

EP 0 644 262 A2

	65	70	75
5	Leu Gly Thr Leu Phe Ile Ile Pro Ala Thr Ser Gln Phe Phe Phe		
	80	85	90
	Asn Ala Leu Pro Ile Leu Thr Trp Val Ala Leu Tyr Phe Thr Ser		
10	95	100	105
	Ser Tyr Phe Pro Asp Asp Arg Arg Pro Pro Ile Thr Val Lys Val		
	110	115	120
15	Leu Pro Ala Val Glu Thr Ile Leu Tyr Gly Asp Asn Leu Ser Asp		
	125	130	135
	Ile Leu Ala Thr Ser Thr Asn Ser Phe Leu Asp Ile Leu Ala Trp		
20	140	145	150
	Leu Pro Tyr Gly Leu Phe His Tyr Gly Ala Pro Phe Val Val Ala		
	155	160	165
25	Ala Ile Leu Phe Val Phe Gly Pro Pro Thr Val Leu Gln Gly Tyr		
	170	175	180
	Ala Phe Ala Phe Gly Tyr Met Asn Leu Phe Gly Val Ile Met Gln		
30	185	190	195
	Asn Val Phe Pro Ala Ala Pro Pro Trp Tyr Lys Ile Leu Tyr Gly		
	200	205	210
35	Leu Gln Ser Ala Asn Tyr Asp Met His Gly Ser Pro Gly Gly Leu		
	215	220	225
	Ala Arg Ile Asp Lys Leu Leu Gly Ile Asn Met Tyr Thr Thr Ala		
40	230	235	240
	Phe Ser Asn Ser Ser Val Ile Phe Gly Ala Phe Pro Ser Leu His		
	245	250	255
45	Ser Gly Cys Ala Thr Met Glu Ala Leu Phe Phe Cys Tyr Cys Phe		
	260	265	270
	Pro Lys Leu Lys Pro Leu Phe Ile Ala Tyr Val Cys Trp Leu Trp		
50	275	280	285
55			

Trp Ser Thr Met Tyr Leu Thr His His Tyr Phe Val Asp Leu Met
 290 295 300
 5 Ala Gly Ser Val Leu Ser Tyr Val Ile Phe Gln Tyr Thr Lys Tyr
 305 310 315
 Thr His Leu Pro Ile Val Asp Thr Ser Leu Phe Cys Arg Trp Ser
 10 320 325 330
 Tyr Thr Ser Ile Glu Lys Tyr Asp Ile Ser Lys Ser Asp Pro Leu
 335 340 345
 15 Ala Ala Asp Ser Asn Asp Ile Glu Ser Val Pro Leu Ser Asn Leu
 350 355 360
 Glu Leu Asp Phe Asp Leu Asn Met Thr Asp Glu Pro Ser Val Ser
 20 365 370 375
 Pro Ser Leu Phe Asp Gly Ser Thr Ser Val Ser Arg Ser Ser Ala
 25 380 385 390
 Thr Ser Ile Thr Ser Leu Gly Val Lys Arg Ala
 395 400

30

SEQ ID NO : 7

SEQUENCE LENGTH : 2340

35

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

40

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

TTTCTTTCTG TCAAAGAATA ATAAAGTGCC CATCAGTGTT CATATTTGTT ACAAAGTGGT 60
 45 TTTCTGATTT GGTACTACTG CAGAGGCGTA TTTTGTGCTT CAGTTACCAT AGCGTAAGAA 120
 CACTAGCGAC TTTTGTTTCGT GAACCAACAG AGTAGGATTT CTACTGCTAC ATCTCTTAGG 180
 TAGTTGGTTA GTCCGATCGC TCACTTTTGG TTGTTGTAA GTACTTCATA AGTTTATCCT 240
 50 TTTCTTTTTT CACACTGAGC TACTTTGGGT ATAGCTTTTG GCCCAAGGAT CTTTGAATTT 300

55

TCTCCAAAAG TACTTTATTT TATATCCTAC AGGTTGCGGT TTTCATATTT TAAAAAGCTT 360
 TTTAATCATT CCTTTGCGTA TGGCAAACCC TTTTTCGAGA TGGTTTCTAT CAGAGAGACC 420
 5 TCCAAACTGC CATGTAGCCG ATTTAGAAAC AAGTTTAGAT CCCCATCAAA CGTTGTTGAA 480
 GGTGCAAAAA TACAAACCCG CTTTAAGCGA CTGGGTGCAT TACATCTTCT TGGGATCCAT 540
 CATGCTGTTT GTGTTCAATTA CTAATCCCGC ACCTTGGATC TTCAAGATCC TTTTTTATTG 600
 10 TTTCTTGGGC ACTTTATTCA TCATTCCAGC TACGTCACAG TTTTCTTCA ATGCCTTGCC 660
 CATCCTAACA TGGGTGGCGC TGTATTTAC TTCATCGTAC TTTCCAGATG ACCGAGGCC 720
 TCCTATTACT GTCAAAGTGT TACCAGCGGT GGAAACAATT TTATACGGCG ACAATTTAAG 780
 15 TGATATTCTT GCAACATCGA CGAATTCCCTT TTTGGACATT TTAGCATGGT TACCGTACGG 840
 ACTATTTTCAT TTTGGGGCCC CATTTGTCGT TGCTGCCATC TTATTCGTAT TTGGTCCACC 900
 20 AACTGTTTTG CAAGGTTATG CTTTTCGATT TGGTTATATG AACCTGTTTG GTGTTATCAT 960
 GCAAAATGTC TTTCCAGCCG CTCCTCCATG GTATAAAATT CTCTATGGAT TGCAATCAGC 1020
 CAACTATGAT ATGCATGGCT CGCCTGGTGG ATTAGCTAGA ATTGATAAGC TACTCGGTAT 1080
 25 TAATATGTAT ACTACAGCTT TTTCAAATTC CTCGTCATT TTCGGTGCTT TTCCTTCACT 1140
 GCATTCCGGG TGTGCTACTA TGAAGCCCT GTTTTCTGT TATTGTTTTC CAAAATTGAA 1200
 GCCCTTGTTT ATTGCTTATG TTTGCTGGT ATGGTGGTCA ACTATGTATC TGACACACCA 1260
 30 TTATTTTGTA GACCTTATGG CAGGTTCTGT GCTGTCATAC GTTATTTTCC AGTACACAAA 1320
 GTACACACAT TTACCAATTG TAGATACATC TCTTTTTTGC AGATGCTCAT AACTTCAAT 1380
 TGAGAAATAC GATATATCAA AGAGTGATCC ATTGGCTGCA GATTCAAACG ATATCGAAAG 1440
 35 TGTCCCTTTG TCCAACCTGG AACTTGACTT TGATCTTAAT ATGACTGATG AACCCAGTGT 1500
 AAGCCCTTCG TTATTTGATG GATCTACTTC TGTCTCTCGT TCGTCCGCCA CGTCTATAAC 1560
 GTCACTAGGT GTAAAGAGGG CTTAATGAGT ATTTTATCTG CAATTACGGA TACGGTTGGT 1620
 40 CTTATGTAGA TACATATAAA TATATATCTT TTTCTTCTT TTTCTTAGTC AGGATTGTCT 1680
 TTTAGCATAA TATACATGTA GTTTATTTAA TCACATACCA CTGATTATCT TTAGAATTTT 1740
 ATAAATTTTT GAAATAAATG GGTGGCTTTT AATGGTGTCT ATGTTAAGTG AGGCTTTTAG 1800
 45 AATGCTCTTC CTGCTTTGTT TATTATATGT GTATGAAAGA TATGTATGTA TTTACATGTG 1860
 TTTGTAGCGT CCCCAGTCAA AACCTGTGCG CTATACCTAA ATGGATTGAT AATCTTCATT 1920
 CACTAATTCT AAAATAGACT TCTCCCCAA AGAACGGTGT AACGATGAGG CTCTATCCAG 1980
 50 CTGCTTATCT AAATCAACTT TAACGATGGA TGATCTTATG ACACGGGGAT CTTTCTTTAA 2040

55

AGTTCTTAGA ATTTCAGACT GTACCGCAGC TGATGAATCA AACAGCATT AAAAGTGATA 2100
 TGCTCGAAAA TGTTTTTCCT GGTCTTTCTT CATTATTTTA GGAAGATACC TTATGCCCAT 2160
 5 GGGTACAATG TCCCTCACCA CACCTCTGTT TTGAATAATC AGTTTCCCGA TTGTGGAAGA 2220
 CAATTCTTTT GCTTCCAAC TTTGGCGCATT GGAGTTGGTT ATGCGAACAA GTCCGATCAG 2280
 CTCATAAAGC ATCTTAGTGA AAAGGGTGGT TTTGCGTTAT TCTTTCCTCT GTTGAAGCTT 2340

SEQ ID NO : 8

SEQUENCE LENGTH : 401

SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Met	Ala	Asn	Pro	Phe	Ser	Arg	Trp	Phe	Leu	Ser	Glu	Arg	Pro	Pro
1				5					10				15	
Asn	Cys	His	Val	Ala	Asp	Leu	Glu	Thr	Ser	Leu	Asp	Pro	His	Gln
				20					25				30	
Thr	Leu	Leu	Lys	Val	Gln	Lys	Tyr	Lys	Pro	Ala	Leu	Ser	Asp	Trp
				35					40				45	
Val	His	Tyr	Ile	Phe	Leu	Gly	Ser	Ile	Met	Leu	Phe	Val	Phe	Ile
				50					55				60	
Thr	Asn	Pro	Ala	Pro	Trp	Ile	Phe	Lys	Ile	Leu	Phe	Tyr	Cys	Phe
				65					70				75	
Leu	Gly	Thr	Leu	Phe	Ile	Ile	Pro	Ala	Thr	Ser	Gln	Phe	Phe	Phe
				80					85				90	
Asn	Ala	Leu	Pro	Ile	Leu	Thr	Trp	Val	Ala	Leu	Tyr	Phe	Thr	Ser
				95					100				105	
Ser	Tyr	Phe	Pro	Asp	Asp	Arg	Arg	Pro	Pro	Ile	Thr	Val	Lys	Val
				110					115				120	

EP 0 644 262 A2

	Leu Pro Ala Val Glu Thr Ile Leu Tyr Gly Asp Asn Leu Ser Asp		
	125	130	135
5	Ile Leu Ala Thr Ser Thr Asn Ser Phe Leu Asp Ile Leu Ala Trp		
	140	145	150
10	Leu Pro Tyr Gly Leu Phe His Phe Gly Ala Pro Phe Val Val Ala		
	155	160	165
	Ala Ile Leu Phe Val Phe Gly Pro Pro Thr Val Leu Gln Gly Tyr		
	170	175	180
15	Ala Phe Ala Phe Gly Tyr Met Asn Leu Phe Gly Val Ile Met Gln		
	185	190	195
20	Asn Val Phe Pro Ala Ala Pro Pro Trp Tyr Lys Ile Leu Tyr Gly		
	200	205	210
	Leu Gln Ser Ala Asn Tyr Asp Met His Gly Ser Pro Gly Gly Leu		
25	215	220	225
	Ala Arg Ile Asp Lys Leu Leu Gly Ile Asn Met Tyr Thr Thr Ala		
	230	235	240
30	Phe Ser Asn Ser Ser Val Ile Phe Gly Ala Phe Pro Ser Leu His		
	245	250	255
	Ser Gly Cys Ala Thr Met Glu Ala Leu Phe Phe Cys Tyr Cys Phe		
35	260	265	270
	Pro Lys Leu Lys Pro Leu Phe Ile Ala Tyr Val Cys Trp Leu Trp		
	275	280	285
40	Trp Ser Thr Met Tyr Leu Thr His His Tyr Phe Val Asp Leu Met		
	290	295	300
	Ala Gly Ser Val Leu Ser Tyr Val Ile Phe Gln Tyr Thr Lys Tyr		
45	305	310	315
	Thr His Leu Pro Ile Val Asp Thr Ser Leu Phe Cys Arg Trp Ser		
	320	325	330
50	Tyr Thr Ser Ile Glu Lys Tyr Asp Ile Ser Lys Ser Asp Pro Leu		

55

335 340 345
 Ala Ala Asp Ser Asn Asp Ile Glu Ser Val Pro Leu Ser Asn Leu
 5 350 355 360
 Glu Leu Asp Phe Asp Leu Asn Met Thr Asp Glu Pro Ser Val Ser
 10 365 370 375
 Pro Ser Leu Phe Asp Gly Ser Thr Ser Val Ser Arg Ser Ser Ala
 380 385 390
 Thr Ser Ile Thr Ser Leu Gly Val Lys Arg Ala
 15 395 400

SEQ ID NO : 9

SEQUENCE LENGTH : 5340

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

AGCGCTTCTA TTTTCCTCCC CACCGCGAGG CGGAAATGGC ACATTTTTTT TCTTTTGCTT 60
 CTGTGCTTTT GCTGTAATTT TTGGCATGTG CTATTGTATG AAGATAACGC GTGGTTCCGT 120
 35 GGAAATAGCC GGAAATTTTG CCGGGAATAT GACGGACATG ATTTAACACC CGTGGAAATG 180
 AAAAAAGCCA AGGTAAGAAA GTGGCAATAT TTTTCCTACA AATAGATCTG CTGTCCCTTA 240
 GATGATTACC ATACATATAT ATATTTATTA CACACATCTG TCAGAGGTAG CTAGCGAAGG 300
 40 TGTCACTGAA ATATTTTTTG TTCCAGTTAG TATAAATACG GAGGTAGAAC AGCTCTCCGC 360
 GTGTATATCT TTTTTTGCGC TATACAAGAA CAGGAAGAAC GCATTTCCAT ACCTTTTTTCT 420
 CCTTACAGGT GCCCTCTGAG TAGTGTACG AACGAGGAAA AAGATTAATA TTAGTGTTTT 480
 45 TATATTCAAA AAGAGTAAAG CCGTTGCTAT ATACGAATAT GACGATTACC GTGGGGGATG 540
 CAGTTTCGGA GACGGAGCTG GAAAACAAAA GTCAAAACGT GGTACTATCT CCCAAGGCAT 600
 CTGCTTCTTC AGACATAAGC ACAGATGTTG ATAAAGACAC ATCGTCTTCT TGGGATGACA 660
 50 AATCTTTGCT GCCTACAGGT GAATATATTG TGGACAGAAA TAAGCCCCAA ACCTACTTGA 720

ATAGCGATGA TATCGAAAAA GTGACAGAAT CTGATATTTT CCCTCAGAAA CGTCTGTTTT 780
 CATTCTTGCA CTCTAAGAAA ATTCCAGAAG TACCACAAAC CGATGACGAG AGGAAGATAT 840
 5 ATCCTCTGTT CCATACAAAT ATTATCTCTA ACATGTTTTT TTGGTGGGTT CTACCCATCC 900
 TGGGAGTTGG TTATAAGAGA ACGATACAGC CGAACGATCT CTTCAAAATG GATCCGAGGA 960
 TGTCTATAGA GACCCTTTAT GACGACTTTG AAAAAAACAT GATTTACTAT TTTGAGAAGA 1020
 10 CGAGGAAAAA ATACCGTAAA AGACATCCAG AAGCGACAGA AGAAGAGGTT ATGGAAAATG 1080
 CCAAACTACC TAAACATACA GTTCTGAGAG CTTTATTATT CACTTTTAAG AAACAGTACT 1140
 TCATGTCGAT AGTGTITGCA ATTCTCGCTA ATTGTACATC CGGTTTTAAC CCCATGATTA 1200
 15 CCAAGAGGCT AATTGAGTTT GTCGAAGAAA AGGCTATTTT TCATAGCATG CATGTTAACA 1260
 AAGGTATTGG TTACGCTATT GGTGCATGTT TGATGATGTT CGTTAACGGG TTGACGTTCA 1320
 ATCATTCTTT TCATACATCC CAACTGACTG GTGTGCAAGC TAAGTCTATT CTTACTAAAG 1380
 20 CTGCCATGAA GAAAATGTTT AATGCATCTA ATTATGCGAG ACATTGTTTT CCTAACGGTA 1440
 AAGTGAATTC TTTTGTAACA ACAGATCTCG CTAGAATTGA ATTTGCCTTA TCTTTTCAGC 1500
 CGTTTTTGGC TGGGTTCCTT GCAATTTTGG CTATTTGCAT TGTTTTATTG ATCGTTAACC 1560
 25 TTGGACCCAT TGCCTTAGTT GGGATTGGTA TTTTTTTCGG TGGGTTTTTC ATATCCTTAT 1620
 TTGCATTTAA GTTAATTCTG GGCTTTAGAA TTGCTGCGAA CATCTTCACT GATGCTAGAG 1680
 TTACCATGAT GAGAGAAGTG CTGAATAATA TAAAAATGAT TAAATATTAT ACGTGGGAGG 1740
 30 ATGCGTATGA AAAAAATATT CAAGATATTA GGACCAAAGA GATTTCTAAA GTTAGAAAAA 1800
 TGCAACTATC AAGAAATTTT TTGATTGCTA TGGCCATGTC TTTGCCTAGT ATTGCTTCAT 1860
 TGGTCACTTT CTTTGCAATG TACAAAGTTA ATAAAGGAGG CAGGCAACCT GGTAAATATT 1920
 35 TTGCCTCTTT ATCTTTATTT CAGGTCTTGA GTTTGCAAAT GTTTTTCTTA CCTATTGCTA 1980
 TTGGTACTGG AATTGACATG ATCATTGGAT TGGGCCGTTT GCAAAGCTTA TTGGAGGCTC 2040
 CAGAAGATGA TCCAAATCAG ATGATTGAAA TGAAGCCCTC TCCTGGCTTT GATCCAAAAT 2100
 40 TGGCTCTAAA AATGACACAT TGCTCATTTG AGTGGGAAGA TTATGAATTA AACGACGCTA 2160
 TTGAAGAAGC AAAAGGAGAA GCTAAAGATG AAGGTAAAAA GAACAAAAAA AAGCGTAAGG 2220
 ATACATGGGG TAAGCCATCT GCAAGTACTA ATAAGGCGAA AAGATTGGAC AATATGTTGA 2280
 45 AAGACAGAGA CGGCCCGGAA GATTTAGAAA AAACCTCGTT TAGGGGTTTC AAGGACTTGA 2340
 ACTTCGATAT TAAAAAGGGC GAATTTATTA TGATTACGGG ACCTATTGGT ACTGGTAAAT 2400
 50 CTTCAATTATT GAATCGCATG GCAGGATCAA TGAGAAAAAT TGATGGTAAG GTTGAAGTCA 2460

55

5
 10
 15
 20
 25
 30
 35
 40
 45
 50
 55

ACGGGGACTT ATTAATGTGT GGTTATCCAT GGATTCAAAA TGCATCTGTA AGAGATAACA 2520
 TCATATTCGG TTCACCATTC AATAAAGAAA AGTATGATGA AGTAGTTCGT GTTTCCTCTT 2580
 TGAAAGCTGA TCTGGATATT TTACCGGCAG GCGATATGAC CGAAATTGGG GAACGTGGTA 2640
 TTACTTTATC TGGTGGTCAA AAGGCACGTA TCAATTTAGC CAGGTCTGTT TATAAGAAGA 2700
 AGGATATTTA TGTATTCGAC GATGTCCTAA GTGCTGTCGA TTCTCGTGTT GGTAAACACA 2760
 TCATGGATGA ATGTCTAACC GGAATGCTTG CTAATAAAAC CAGAATTTTA GCAACGCATC 2820
 AGTTGTCACT GATTGAGAGA GCTTCTAGAG TCATCGTTTT AGGTACTGAT GGCCAAGTCG 2880
 ATATTGGTAC TGTTGATGAG CTAAGAGCTC GTAATCAAAC TTTGATAAAT CTTTTACAAT 2940
 TCTCTTCTCA AAATTCGGAG AAAGAGGATG AAGAACAGGA AGCGGTTGTT TCCGGTGAAT 3000
 TGGGACAACT AAAATATGAA CCAGAGGTAA AGGAATTGAC TGAAGTGAAG AAAAAGGCTA 3060
 CAGAAATGTC ACAAAGTCA AATAGTGGTA AAATTGTAGC GGATGGTCAT ACTAGTAGTA 3120
 AAGAAGAAAG AGCAGTCAAT AGTATCAGTC TGAAGATATA CCGTGAATAC ATTAAAGCTG 3180
 CAGTAGGTAA GTGGGGTTTT ATCGCACTAC CGTTGTATGC AATTTTAGTC GTTGGAACCA 3240
 CATTCTGCTC ACTTTTTTCT TCCGTTTGGT TATCTTACTG GACTGAGAAT AAATTCAAAA 3300
 ACAGACCACC CAGTTTTTAT ATGGGTCTTT ACTCCTTCTT TGTGTTTGCT GCTTTCATAT 3360
 TCATGAATGG CCAGTTCACC ATACTTTGCG CAATGGGTAT TATGGCATCG AAATGGTTAA 3420
 ATTTGAGGGC TGTGAAAAGA ATTTTACACA CTCCAATGTC ATACATAGAT ACCACACCTT 3480
 TGGGACGTAT TCTGAACAGA TTCACAAAAG ATACAGATAG CTTAGATAAT GAGTTAACCG 3540
 AAAGTTTACG GTTGATGACA TCTCAATTG CTAATATTGT AGGTGTTTGC GTCATGTGTA 3600
 TTGTTTACTT GCCGTGGTTT GCTATCGCAA TTCCGTTTCT TTTGGTCATC TTTGTTCTGA 3660
 TTGCTGATCA TTATCAGAGT TCTGGTAGAG AAATTAAAAG ACTTGAAGCT GTGCAACGGT 3720
 CTTTTGTTTA CAATAATTTA AATGAAGTTT TGGGTGGGAT GGATACAATC AAAGCATACC 3780
 GAAGTCAGGA ACGATTTTGT GCGAAATCAG ATTTTGTGAT CAACAAGATG AATGAGGCGG 3840
 GATACCTTGT AGTTGTCCTG CAAAGATGGG TAGGTATTTT CCTTGATATG GTTGCTATCG 3900
 CATTTGCACT AATTATTACG TTATTGTGTG TTACGAGAGC CTTTCCTATT TCCGCGGCTT 3960
 CAGTTGGTGT TTTGTTGACT TATGTATTAC AATTGCCTGG TCTATTAAAT ACCATTTTAA 4020
 GGGCAATGAC TCAAACAGAG AATGACATGA ATAGTCCGA AAGATTGGTA ACATATGCAA 4080
 CTGAACTACC ACTAGAGGCA TCCTATAGAA AGCCCGAAAT GACACCTCCA GAGTCATGGC 4140
 CCTCAATGGG CGAAATAATT TTTGAAAATG TTGATTTTGC CTATAGACCT GGTTTACCTA 4200

TAGTTTTAAA AAATCTTAAC TTGAATATCA AGAGTGGGGA AAAAATTGGT ATCTGTGGTC 4260
 GTACAGGTGC TGGTAAGTCC ACTATTATGA GTGCCCTTTA CAGGTTGAAT GAATTGACCG 4320
 CAGGTAAAAAT TTTAATTGAC AATGTTGATA TAAGTCAGCT GGGACTTTTC GATTTAAGAA 4380
 GAAAATTAGC CATCATTCCA CAAGATCCAG TATTATTTAG GGGTACGATT CGCAAGAACT 4440
 TAGATCCATT TAATGAGCGT ACAGATGACG AATTATGGGA TGCATTGGTG AGAGGTGGTG 4500
 CTATCGCCAA GGATGACTTG CCGGAAGTGA AATTGCAAAA ACCTGATGAA AATGGTACTC 4560
 ATGGTAAAAAT GCATAAGTTC CATTTAGATC AAGCAGTGGG AGAAGAGGGC TCCAATTTCT 4620
 CCTTAGGTGA GAGACAACTA TTAGCATTAA CAAGGGCATT GGTCCGCCAA TCAAAAATAT 4680
 TGATTTTGGG TGAGGCTACA TCCTCAGTGG ACTACGAAAC GGATGGCAAA ATCCAAACAC 4740
 GTATTGTTGA GGAATTTGGA GATTGTACAA TTTTGTGTAT TGCTCACAGA CTGAAGACCA 4800
 TTGTAAATTA TGATCGTATT CTTGTTTTAG AGAAGGGTGA AGTCGCAGAA TTCGATACAC 4860
 CATGGACGTT GTTTAGTCAA GAAGATAGTA TTTTCAGAAG CATGTGTTCT AGATCTGGTA 4920
 TTGTGAAAAA TGATTTGAG AACAGAAATT AATTTATATT ATTTGTTGCA TGATTTTCT 4980
 CTTTTATTTA TTTATATGTT GCCGATGGTA CAAATTAGTA CTAGAAAAGA AAACCCACTA 5040
 CTATGACTTG CAGAAAAAGT TATGTGTGCC ATAGATAGAT ATAATTGCAT ACCCACATCG 5100
 TATACTCAAA ATTCCGAAAA GAACATTTCA TTTTTTATGA GGCAAACTGA ACAACGCTTC 5160
 GGTCTTTTTT TCATTCTAGA AATATATATT TATACATCAT TTTCAGAAGA TATTCAAAGA 5220
 ACTTATTGGG ATGTCTATTT ACTGAATAAA GTATACACAA AAAACGAATT TAAAATGGAA 5280
 GGCATAAATA GAAAACCTAG AAGTGAAAAT CCTAAAACCG AAGGATATTT CAAATACGTA 5340

SEQ ID NO : 10

SEQUENCE LENGTH : 1477

SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Met Thr Ile Thr Val Gly Asp Ala Val Ser Glu Thr Glu Leu Glu

5

10

15

EP 0 644 262 A2

	Asn Lys Ser Gln Asn Val Val Leu Ser Pro Lys Ala Ser Ala Ser	
	20	25 30
5	Ser Asp Ile Ser Thr Asp Val Asp Lys Asp Thr Ser Ser Ser Trp	
	35	40 45
10	Asp Asp Lys Ser Leu Leu Pro Thr Gly Glu Tyr Ile Val Asp Arg	
	50	55 60
	Asn Lys Pro Gln Thr Tyr Leu Asn Ser Asp Asp Ile Glu Lys Val	
15	65	70 75
	Thr Glu Ser Asp Ile Phe Pro Gln Lys Arg Leu Phe Ser Phe Leu	
	80	85 90
20	His Ser Lys Lys Ile Pro Glu Val Pro Gln Thr Asp Asp Glu Arg	
	95	100 105
	Lys Ile Tyr Pro Leu Phe His Thr Asn Ile Ile Ser Asn Met Phe	
25	110	115 120
	Phe Trp Trp Val Leu Pro Ile Leu Arg Val Gly Tyr Lys Arg Thr	
	125	130 135
30	Ile Gln Pro Asn Asp Leu Phe Lys Met Asp Pro Arg Met Ser Ile	
	140	145 150
	Glu Thr Leu Tyr Asp Asp Phe Glu Lys Asn Met Ile Tyr Tyr Phe	
35	155	160 165
	Glu Lys Thr Arg Lys Lys Tyr Arg Lys Arg His Pro Glu Ala Thr	
	170	175 180
40	Glu Glu Glu Val Met Glu Asn Ala Lys Leu Pro Lys His Thr Val	
	185	190 195
	Leu Arg Ala Leu Leu Phe Thr Phe Lys Lys Gln Tyr Phe Met Ser	
45	200	205 210
	Ile Val Phe Ala Ile Leu Ala Asn Cys Thr Ser Gly Phe Asn Pro	
50	215	220 225
	Met Ile Thr Lys Arg Leu Ile Glu Phe Val Glu Glu Lys Ala Ile	

55

EP 0 644 262 A2

	230	235	240
	Phe His Ser Met His Val Asn Lys Gly Ile Gly Tyr Ala Ile Gly		
5	245	250	255
	Ala Cys Leu Met Met Phe Val Asn Gly Leu Thr Phe Asn His Phe		
	260	265	270
10	Phe His Thr Ser Gln Leu Thr Gly Val Gln Ala Lys Ser Ile Leu		
	275	280	285
	Thr Lys Ala Ala Met Lys Lys Met Phe Asn Ala Ser Asn Tyr Ala		
15	290	295	300
	Arg His Cys Phe Pro Asn Gly Lys Val Thr Ser Phe Val Thr Thr		
	305	310	315
20	Asp Leu Ala Arg Ile Glu Phe Ala Leu Ser Phe Gln Pro Phe Leu		
	320	325	330
	Ala Gly Phe Pro Ala Ile Leu Ala Ile Cys Ile Val Leu Leu Ile		
25	335	340	345
	Val Asn Leu Gly Pro Ile Ala Leu Val Gly Ile Gly Ile Phe Phe		
	350	355	360
30	Gly Gly Phe Phe Ile Ser Leu Phe Ala Phe Lys Leu Ile Leu Gly		
	365	370	375
	Phe Arg Ile Ala Ala Asn Ile Phe Thr Asp Ala Arg Val Thr Met		
35	380	385	390
	Met Arg Glu Val Leu Asn Asn Ile Lys Met Ile Lys Tyr Tyr Thr		
40	395	400	405
	Trp Glu Asp Ala Tyr Glu Lys Asn Ile Gln Asp Ile Arg Thr Lys		
	410	415	420
45	Glu Ile Ser Lys Val Arg Lys Met Gln Leu Ser Arg Asn Phe Leu		
	425	430	435
	Ile Ala Met Ala Met Ser Leu Pro Ser Ile Ala Ser Leu Val Thr		
50	440	445	450
55			

EP 0 644 262 A2

	Phe	Leu	Ala	Met	Tyr	Lys	Val	Asn	Lys	Gly	Gly	Arg	Gln	Pro	Gly
						455				460					465
5	Asn	Ile	Phe	Ala	Ser	Leu	Ser	Leu	Phe	Gln	Val	Leu	Ser	Leu	Gln
						470				475					480
10	Met	Phe	Phe	Leu	Pro	Ile	Ala	Ile	Gly	Thr	Gly	Ile	Asp	Met	Ile
						485				490					495
	Ile	Gly	Leu	Gly	Arg	Leu	Gln	Ser	Leu	Leu	Glu	Ala	Pro	Glu	Asp
						500				505					510
15	Asp	Pro	Asn	Gln	Met	Ile	Glu	Met	Lys	Pro	Ser	Pro	Gly	Phe	Asp
						515				520					525
20	Pro	Lys	Leu	Ala	Leu	Lys	Met	Thr	His	Cys	Ser	Phe	Glu	Trp	Glu
						530				535					540
	Asp	Tyr	Glu	Leu	Asn	Asp	Ala	Ile	Glu	Glu	Ala	Lys	Gly	Glu	Ala
25						545				550					555
	Lys	Asp	Glu	Gly	Lys	Lys	Asn	Lys	Lys	Lys	Arg	Lys	Asp	Thr	Trp
						560				565					570
30	Gly	Lys	Pro	Ser	Ala	Ser	Thr	Asn	Lys	Ala	Lys	Arg	Leu	Asp	Asn
						575				580					585
	Met	Leu	Lys	Asp	Arg	Asp	Gly	Pro	Glu	Asp	Leu	Glu	Lys	Thr	Ser
35						590				595					600
	Phe	Arg	Gly	Phe	Lys	Asp	Leu	Asn	Phe	Asp	Ile	Lys	Lys	Gly	Glu
						605				610					615
40	Phe	Ile	Met	Ile	Thr	Gly	Pro	Ile	Gly	Thr	Gly	Lys	Ser	Ser	Leu
						620				625					630
	Leu	Asn	Ala	Met	Ala	Gly	Ser	Met	Arg	Lys	Ile	Asp	Gly	Lys	Val
45						635				640					645
	Glu	Val	Asn	Gly	Asp	Leu	Leu	Met	Cys	Gly	Tyr	Pro	Trp	Ile	Gln
						650				655					660
50	Asn	Ala	Ser	Val	Arg	Asp	Asn	Ile	Ile	Phe	Gly	Ser	Pro	Phe	Asn

55

EP 0 644 262 A2

	665	670	675
5	Lys Glu Lys Tyr Asp Glu Val Val Arg Val Cys Ser Leu Lys Ala		
	680	685	690
	Asp Leu Asp Ile Leu Pro Ala Gly Asp Met Thr Glu Ile Gly Glu		
10	695	700	705
	Arg Gly Ile Thr Leu Ser Gly Gly Gln Lys Ala Arg Ile Asn Leu		
	710	715	720
15	Ala Arg Ser Val Tyr Lys Lys Lys Asp Ile Tyr Val Phe Asp Asp		
	725	730	735
	Val Leu Ser Ala Val Asp Ser Arg Val Gly Lys His Ile Met Asp		
20	740	745	750
	Glu Cys Leu Thr Gly Met Leu Ala Asn Lys Thr Arg Ile Leu Ala		
	755	760	765
25	Thr His Gln Leu Ser Leu Ile Glu Arg Ala Ser Arg Val Ile Val		
	770	775	780
	Leu Gly Thr Asp Gly Gln Val Asp Ile Gly Thr Val Asp Glu Leu		
30	785	790	795
	Lys Ala Arg Asn Gln Thr Leu Ile Asn Leu Leu Gln Phe Ser Ser		
	800	805	810
35	Gln Asn Ser Glu Lys Glu Asp Glu Glu Gln Glu Ala Val Val Ser		
	815	820	825
	Gly Glu Leu Gly Gln Leu Lys Tyr Glu Pro Glu Val Lys Glu Leu		
40	830	835	840
	Thr Glu Leu Lys Lys Lys Ala Thr Glu Met Ser Gln Thr Ala Asn		
	845	850	855
45	Ser Gly Lys Ile Val Ala Asp Gly His Thr Ser Ser Lys Glu Glu		
	860	865	870
	Arg Ala Val Asn Ser Ile Ser Leu Lys Ile Tyr Arg Glu Tyr Ile		
50	875	880	885

55

EP 0 644 262 A2

	Lys Ala Ala Val Gly Lys Trp Gly Phe Ile Ala Leu Pro Leu Tyr		
	890	895	900
5	Ala Ile Leu Val Val Gly Thr Thr Phe Cys Ser Leu Phe Ser Ser		
	905	910	915
10	Val Trp Leu Ser Tyr Trp Thr Glu Asn Lys Phe Lys Asn Arg Pro		
	920	925	930
	Pro Ser Phe Tyr Met Gly Leu Tyr Ser Phe Phe Val Phe Ala Ala		
15	935	940	945
	Phe Ile Phe Met Asn Gly Gln Phe Thr Ile Leu Cys Ala Met Gly		
	950	955	960
20	Ile Met Ala Ser Lys Trp Leu Asn Leu Arg Ala Val Lys Arg Ile		
	965	970	975
	Leu His Thr Pro Met Ser Tyr Ile Asp Thr Thr Pro Leu Gly Arg		
25	980	985	990
	Ile Leu Asn Arg Phe Thr Lys Asp Thr Asp Ser Leu Asp Asn Glu		
	995	1000	1005
30	Leu Thr Glu Ser Leu Arg Leu Met Thr Ser Gln Phe Ala Asn Ile		
	1010	1015	1020
	Val Gly Val Cys Val Met Cys Ile Val Tyr Leu Pro Trp Phe Ala		
35	1025	1030	1035
	Ile Ala Ile Pro Phe Leu Leu Val Ile Phe Val Leu Ile Ala Asp		
	1040	1045	1050
40	His Tyr Gln Ser Ser Gly Arg Glu Ile Lys Arg Leu Glu Ala Val		
	1055	1060	1065
	Gln Arg Ser Phe Val Tyr Asn Asn Leu Asn Glu Val Leu Gly Gly		
45	1070	1075	1080
	Met Asp Thr Ile Lys Ala Tyr Arg Ser Gln Glu Arg Phe Leu Ala		
50	1085	1090	1095
	Lys Ser Asp Phe Leu Ile Asn Lys Met Asn Glu Ala Gly Tyr Leu		

55

EP 0 644 262 A2

	1100	1105	1110
	Val Val Val Leu Gln Arg Trp Val Gly Ile Phe Leu Asp Met Val		
5	1115	1120	1125
	Ala Ile Ala Phe Ala Leu Ile Ile Thr Leu Leu Cys Val Thr Arg		
10	1130	1135	1140
	Ala Phe Pro Ile Ser Ala Ala Ser Val Gly Val Leu Leu Thr Tyr		
	1145	1150	1155
15	Val Leu Gln Leu Pro Gly Leu Leu Asn Thr Ile Leu Arg Ala Met		
	1160	1165	1170
	Thr Gln Thr Glu Asn Asp Met Asn Ser Ala Glu Arg Leu Val Thr		
20	1175	1180	1185
	Tyr Ala Thr Glu Leu Pro Leu Glu Ala Ser Tyr Arg Lys Pro Glu		
	1190	1195	1200
25	Met Thr Pro Pro Glu Ser Trp Pro Ser Met Gly Glu Ile Ile Phe		
	1205	1210	1215
	Glu Asn Val Asp Phe Ala Tyr Arg Pro Gly Leu Pro Ile Val Leu		
30	1220	1225	1230
	Lys Asn Leu Asn Leu Asn Ile Lys Ser Gly Glu Lys Ile Gly Ile		
	1235	1240	1245
35	Cys Gly Arg Thr Gly Ala Gly Lys Ser Thr Ile Met Ser Ala Leu		
	1250	1255	1260
	Tyr Arg Leu Asn Glu Leu Thr Ala Gly Lys Ile Leu Ile Asp Asn		
40	1265	1270	1275
	Val Asp Ile Ser Gln Leu Gly Leu Phe Asp Leu Arg Arg Lys Leu		
	1280	1285	1290
45	Ala Ile Ile Pro Gln Asp Pro Val Leu Phe Arg Gly Thr Ile Arg		
	1295	1300	1305
	Lys Asn Leu Asp Pro Phe Asn Glu Arg Thr Asp Asp Glu Leu Trp		
50	1310	1315	1320

55

EP 0 644 262 A2

	Asp Ala Leu Val Arg Gly Gly Ala Ile Ala Lys Asp Asp Leu Pro		
	1325	1330	1335
5	Glu Val Lys Leu Gln Lys Pro Asp Glu Asn Gly Thr His Gly Lys		
	1340	1345	1350
	Met His Lys Phe His Leu Asp Gln Ala Val Glu Glu Glu Gly Ser		
10	1355	1360	1365
	Asn Phe Ser Leu Gly Glu Arg Gln Leu Leu Ala Leu Thr Arg Ala		
	1370	1375	1380
15	Leu Val Arg Gln Ser Lys Ile Leu Ile Leu Asp Glu Ala Thr Ser		
	1385	1390	1395
	Ser Val Asp Tyr Glu Thr Asp Gly Lys Ile Gln Thr Arg Ile Val		
20	1400	1405	1410
	Glu Glu Phe Gly Asp Cys Thr Ile Leu Cys Ile Ala His Arg Leu		
	1415	1420	1425
25	Lys Thr Ile Val Asn Tyr Asp Arg Ile Leu Val Leu Glu Lys Gly		
	1430	1435	1440
30	Glu Val Ala Glu Phe Asp Thr Pro Trp Thr Leu Phe Ser Gln Glu		
	1445	1450	1455
	Asp Ser Ile Phe Arg Ser Met Cys Ser Arg Ser Gly Ile Val Glu		
35	1460	1465	1470
	Asn Asp Phe Glu Asn Arg Ser		
	1475		

40	SEQ ID NO : 11
	SEQUENCE LENGTH : 26
	SEQUENCE TYPE : nucleic acid
45	STRANDEDNESS : single
	TOPOLOGY : linear
	MOLECULE TYPE : Other nucleic acid(synthetic DNA)
50	SEQUENCE DESCRIPTION :

55

TTTGTTTAYA TGAAYTTNTT YGGNGT 26

5

SEQ ID NO : 12

SEQUENCE LENGTH : 29

SEQUENCE TYPE : nucleic acid

10

STRANDEDNESS : single

TOPOLOGY : linear

MOLECULE TYPE : Other nucleic acid(synthetic DNA)

15

SEQUENCE DESCRIPTION :

TCTACAAART ARTGGTGNGT NARRTACAT 29

20

SEQ ID NO : 13

SEQUENCE LENGTH : 2274

25

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

30

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

TTATATATAT TATTGATTTG TTCCTGTTGT TATTTAGTTT AGAATCAGAC GACTACACCA 60

35

GAACCACAAT TCAACCAACA CTTATATAGA ACCTGGCTTG GAAAAAAGTA ACATTTATCA 120

TTCCTATACT TTTTGTAGCAA ACATAATCCG TGTGTTTACAT ATATTATTCA CCCAATATCA 180

TAACAAAAAC AAAGTGAATA ATGGCGTCTT CTATTTTGCG TTCCAAAATA ATACAAAAAC 240

40

CGTACCAATT ATTCCACTAC TATTTTCTTC TGGAGAAGGC TCCTGGTTCT ACAGTTAGTG 300

ATTTGAATTT TGATACAAAC ATACAAACGA GTTTACGTAA ATTAAAGCAT CATCATTGGA 360

CGGTGGGAGA AATATTCCAT TATGGGTTTT TGGTTTCCAT ACTTTTTTTC GTGTTTGTGG 420

45

TTTTCCCAGC TTCATTTTTT ATAAATTAC CAATAATCTT AGCATTGCT ACTTGTTTTT 480

TAATACCCTT AACATCACAA TTTTCTTCTC CTGCCTTGCC CGTTTCACT TGGTTGGCAT 540

TATATTTTAC GTGTGCTAAA ATACCTCAAG AATGGAAACC AGCTATCACA GTTAAAGTTT 600

50

TACCAGCTAT GGAAACAATT TTGTACGGCG ATAATTTATC AAATGTTTTG GCAACCATCA 660

55

CTACCGGAGT GTTAGATATA TTGGCATGGT TACCATATGG GATTATTCAT TTCAGTTTCC 720
CATTGTACT TGCTGCTATT ATATTTTAT TTGGGCCACC GACGGCATTAGATCATTG 780
GATTGCGCTT TGGTTATATG AACTTGCTTG GAGTCTTGAT TCAAATGGCA TTCCCAGCTG 840
CTCCTCCATG GTACAAAAAC TTGCACGGAT TAGAACCAGC TAATTATTCA ATGCACGGGT 900
CTCCTGGTGG ACTTGAAGG ATAGATAAAT TGTTAGGTGT TGATATGTAT ACCACAGGGT 960
TTTCCAATTC ATCAATCATT TTTGGGGCAT TCCCATCGTT ACATTCAGGA TGTTGTATCA 1020
TGGAAGTGTT ATTTTGTGT TGGTTGTTTC CACGATTCAA GTTTGTGTGG GTTACATACG 1080
CATCTGGCT TTGGTGGAGC ACGATGTATT TGACCCATCA CTACTTTGTC GATTTGATTG 1140
GTGGAGCCAT GCTATCTTTG ACTGTTTTTG AGTTCACCAA ATATAAATAT TTGCCAAAAA 1200
ACAAAGAAGG CCTTTTCTGT CGTTGGTCAT AACTGAAAT TGAAAAAATC GATATCCAAG 1260
AGATTGACCC TTTATCATAC AATTATATCC CTGTCAACAG CAATGATAAT GAAAGCAGAT 1320
TGTATACGAG AGTGTAACAA GAGTCTCAGG TTAGTCCCCC ACAGAGAGCT GAAACACCTG 1380
AAGCATTGA GATGTCAAAT TTTTCTAGGT CTAGACAAAG CTCAAAGACT CAGGTTCCAT 1440
TGAGTAATCT TACTAACAAT GATCAAGTGT CTGGAATTAA CGAAGAGGAT GAAGAAGAAG 1500
AAGGCGATGA AATTTTCATCG AGTACTCCTT CGGTGTTTGA AGACGAACCA CAGGGTAGCA 1560
CATATGCTGC ATCCTCAGCT ACATCAGTAG ATGATTTGGA TTCCAAAAGA AATTAGTAAA 1620
ATAACAGTTT CTATTAATTT CTTTATTTCC TCCTAATTAA TGATTTTATG CTCAATACCT 1680
ACACTATCTG TTTTAAATTT CCTACTTTTT TTTTATTATT GTTGAGTTCA TTTGCTGTTC 1740
ATTGAATATT TACAATTTTG CATTAATTAC CATCAATATA GAATGGGCAC AGTTTTTTTA 1800
AGTTTTTTTG TTTTGTGTT TGTCTTCTT TTTTACATT AATGTGTTG GATTGTTTTA 1860
GGTTCCTTTA TCCCTTAGCC CCCTCAGAAT ACTATTTTAT CTAATTAATT TGTTTTTATT 1920
TTCTGATATT TACCAATTGC TTTTCTTTT GGATATTTAT AATAGCATCC CCTAATAATT 1980
AATATACAAC TGTTTCATAT ATATACGTGT ATGTCCTGTA GTGGTGAAA CTGGAGTCAA 2040
CATTGTATT AATGTGTACA AGAAAGCAGT GTTAATGCTA CTATTATAAT TTTTGAGGTG 2100
CAAATCAAGA GGTTGGCAGC TTTCTTATGG CTATGACCGT GAATGAAGGC TTGTAAACCA 2160
CGTAATAAAC AAAAGCCAAC AAGTTTTTTT AGAGCCTTTA ACAACATACG CAATGAGAGT 2220
GATTGCAATA CTACAAGATA TAGCCCAAAA AATTGAATGC ATTTCAACAA CAAC 2274

SEQ ID NO : 14

SEQUENCE LENGTH : 471

SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Met Ala Ser Ser Ile Leu Arg Ser Lys Ile Ile Gln Lys Pro Tyr

5 10 15

Gln Leu Phe His Tyr Tyr Phe Leu Ser Glu Lys Ala Pro Gly Ser

20 25 30

Thr Val Ser Asp Leu Asn Phe Asp Thr Asn Ile Gln Thr Ser Leu

35 40 45

Arg Lys Leu Lys His His His Trp Thr Val Gly Glu Ile Phe His

50 55 60

Tyr Gly Phe Leu Val Ser Ile Leu Phe Phe Val Phe Val Val Phe

65 70 75

Pro Ala Ser Phe Phe Ile Lys Leu Pro Ile Ile Leu Ala Phe Ala

80 85 90

Thr Cys Phe Leu Ile Pro Leu Thr Ser Gln Phe Phe Leu Pro Ala

95 100 105

Leu Pro Val Phe Thr Trp Leu Ala Leu Tyr Phe Thr Cys Ala Lys

110 115 120

Ile Pro Gln Glu Trp Lys Pro Ala Ile Thr Val Lys Val Leu Pro

125 130 135

Ala Met Glu Thr Ile Leu Tyr Gly Asp Asn Leu Ser Asn Val Leu

140 145 150

Ala Thr Ile Thr Thr Gly Val Leu Asp Ile Leu Ala Trp Leu Pro

155 160 165

Tyr Gly Ile Ile His Phe Ser Phe Pro Phe Val Leu Ala Ala Ile

EP 0 644 262 A2

	170	175	180
	Ile Phe Leu Phe Gly Pro Pro Thr Ala Leu Arg Ser Phe Gly Phe		
5	185	190	195
	Ala Phe Gly Tyr Met Asn Leu Leu Gly Val Leu Ile Gln Met Ala		
10	200	205	210
	Phe Pro Ala Ala Pro Pro Trp Tyr Lys Asn Leu His Gly Leu Glu		
	215	220	225
15	Pro Ala Asn Tyr Ser Met His Gly Ser Pro Gly Gly Leu Gly Arg		
	230	235	240
	Ile Asp Lys Leu Leu Gly Val Asp Met Tyr Thr Thr Gly Phe Ser		
20	245	250	255
	Asn Ser Ser Ile Ile Phe Gly Ala Phe Pro Ser Leu His Ser Gly		
	260	265	270
25	Cys Cys Ile Met Glu Val Leu Phe Leu Cys Trp Leu Phe Pro Arg		
	275	280	285
	Phe Lys Phe Val Trp Val Thr Tyr Ala Ser Trp Leu Trp Trp Ser		
30	290	295	300
	Thr Met Tyr Leu Thr His His Tyr Phe Val Asp Leu Ile Gly Gly		
	305	310	315
35	Ala Met Leu Ser Leu Thr Val Phe Glu Phe Thr Lys Tyr Lys Tyr		
	320	325	330
	Leu Pro Lys Asn Lys Glu Gly Leu Phe Cys Arg Trp Ser Tyr Thr		
40	335	340	345
	Glu Ile Glu Lys Ile Asp Ile Gln Glu Ile Asp Pro Leu Ser Tyr		
	350	355	360
45	Asn Tyr Ile Pro Val Asn Ser Asn Asp Asn Glu Ser Arg Leu Tyr		
	365	370	375
50	Thr Arg Val Tyr Gln Glu Ser Gln Val Ser Pro Pro Gln Arg Ala		
	380	385	390

55

Glu Thr Pro Glu Ala Phe Glu Met Ser Asn Phe Ser Arg Ser Arg
 395 400 405
 5 Gln Ser Ser Lys Thr Gln Val Pro Leu Ser Asn Leu Thr Asn Asn
 410 415 420
 10 Asp Gln Val Ser Gly Ile Asn Glu Glu Asp Glu Glu Glu Gly
 425 430 435
 Asp Glu Ile Ser Ser Ser Thr Pro Ser Val Phe Glu Asp Glu Pro
 440 445 450
 15 Gln Gly Ser Thr Tyr Ala Ala Ser Ser Ala Thr Ser Val Asp Asp
 455 460 465
 20 Leu Asp Ser Lys Arg Asn
 470

SEQ ID NO : 15

25 SEQUENCE LENGTH : 243

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

30 TOPOLOGY : linear

MOLECULE TYPE : genomic DNA

SEQUENCE DESCRIPTION :

35 TTTGAAAAAT TTGAATTTTA AAATTAATCC AATGGAAAAA ATTGGTATTT GTGGAAGAAC 60
 CGGTGCTGGT AAATCATCAA TTATGACAGC ATTATATCGA TTATCAGAAT TAGAACTGGG 120
 GAAAATTATT ATTGATGATA TTGATATTTT AACTTTGGGT TAAAAGATC TTCGATCAAA 180
 40 ATTATCAATT ATTCCTCAAG ATCCAGTATT ATTCCGAGGT TCAATTCGGA AAAACTTGGA 240
 TCC 243

45 SEQ ID NO : 16

SEQUENCE LENGTH : 80

50 SEQUENCE TYPE : amino acid

STRANDEDNESS : single

55

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Leu Lys Asn Leu Asn Phe Lys Ile Asn Pro Met Glu Lys Ile Gly

5 10 15

Ile Cys Gly Arg Thr Gly Ala Gly Lys Ser Ser Ile Met Thr Ala

20 25 30

Leu Tyr Arg Leu Ser Glu Leu Glu Leu Gly Lys Ile Ile Ile Asp

35 40 45

Asp Ile Asp Ile Ser Thr Leu Gly Leu Lys Asp Leu Arg Ser Lys

50 55 60

Leu Ser Ile Ile Pro Gln Asp Pro Val Leu Phe Arg Gly Ser Ile

65 70 75

Arg Lys Asn Leu Asp

80

SEQ ID NO : 17

SEQUENCE LENGTH : 1601

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

MOLECULE TYPE : genomic DNA

ANTI-SENSE : Yes

SEQUENCE DESCRIPTION :

AGGAAGATGA CTTGCATCAA AGATGGAGGA AGTGGTACTG GCAGGACGAT CAATCAAATC 60

AGCAGCAGGA CTAGGTAACG GCTCAGGTGA TGATGAACCC ACGGACCATT CATGATCGGT 120

GTTAGCAAGT TCCATATTGT TAAGACCACT CATGAAGGCT ACTGCATTAG GGTTTTGAGT 180

AAAAGAATCC CTTCCAAGTA AGTATGGGCT GCCGGTACGA GCCAAGGAGT TGCTGGTTTT 240

TTCGGAAAGA CCATGACCGT GGATAACAAA CTCGTATTCC CAACGAAGGA TTTTACCAGT 300

TTGCAACTGT GGGAGGCGTA GCTTTTGAGC AAAAACGAAG CATATAATAG CTAAACACAT 360

ACGCGCGACC AAATCTACAA AGTAGTGGTG GGTAAGGTAC ATAGTACACC AGCAAAGCCA 420
TAGAACATAT CCATAAAAGC AGAAGCGGTA TCGAGGAAAC ACATGCGAAA GGAAAAGTGC 480
5 TTCCAGCATG GCCCATCCAG CGTGAAGAGA TGGAAAGGCA CCAAAAACAA CCGGAGAGTT 540
AGAAAAACCA TCAGTGTAAG TGCTAGTGGC GAAGAGAGCA TCAATACGGG CCAATCCACC 600
AGGAGAGCCA CGTACTGCAT ACGTGGCAGG TTCTAAACCA TACATATTTT CATACCAAGG 660
10 AGGAGAACAG GGGAAAGCCA TTTGGATAAG AACACCAAAT AAATTCATAT AACCAAAAGT 720
TCGAGCCCAA ACTGGAAGAG TTCCAGGAGG TGCAAAGATG AAAAGAATAA ATGAAATGAT 780
AAAAGGAGCC GAATAATGCA TGAATCCATA TGGAAACCAG GCCAAAATAT CAAGGATGCT 840
15 ATGCGTGGTT TTCGAGAGAA GACTAGAAAG ATTAGAGCCA TAAAGAATAT TTTCAAGTGT 900
GGGTAAACAA CGAACCATA TGGGTGGACG CCAGCGTTCT GGAATAAACC TACAAGAGTA 960
20 AAATAAAATT GCCCAGGTGA TGATAACAAT GGCAGGAAAA AAAATTTGGC GTGTTAAAGG 1020
AACGGTCAAC GCAATGGCCA AAAGACAGGC AATGCCAAAT TTCCCCAGA ATCCAGGAGA 1080
TTCAATGACA ATACAAGCAA AAATCAAATT ACCTGCTAGA AACACATATT GCAAATGTGT 1140
25 CCATGACCAT TTCGTATTGC GTAGCAAACG AAATGTAGGC ATAGGGTTTA AGCTTGTTTC 1200
CAACTTGTAT TGGGATGCTC GGTTACACGC AGCAAGGCGC TTTTTTAAGG TCGAAAGAGC 1260
AGACATTGCT TCAAAGAATT ATCAGAGTAA AAAAGGGAAG CGTACGAAAA AAATTTCTGT 1320
30 AGGAATTAAC CGGAAAACCTA AAGGAAAAAA AAGGAATTTT TATGAAGGAA AGAAAGTAGC 1380
TATTAAATGC AAGTGTCAAG CACTTAAAAG TAGCGATGTA AAATATTTAA AAAAAGATGG 1440
ACCGATTAAC CAATGTTTCTAG CTCACAGTTG CCAGCAATCA GGGCTATTTT TTTATTTTTT 1500
35 TTATAAAATT GCTAATTATA TATAATATAA TTAGTTTATT AACTTGCTTT TCCTCAAAAA 1560
ACCAATTCTG GAAAGGAACT TTTGCAGAGG CAAAAAAGCT T 1601

40 SEQ ID NO : 18

SEQUENCE LENGTH : 1601

45 SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

50 MOLECULE TYPE : mRNA

ANTI-SENSE : Yes

SEQUENCE DESCRIPTION :

5 AGGAAGAUGA CUUGCAUCAA AGAUGGAGGA AGUGGUACUG GCAGGACGAU CAAUCAAUUC 60
 AGCAGCAGGA CUAGGUAACG GCUCAGGUGA UGAUGAACCC ACGGACCAUU CAUGAUCGGU 120
 GUUAGCAAGU UCCAUUUUGU UAAGACCACU CAUGAAGGCU ACUGCAUUAG GGUUUUGAGU 180
 10 AAAAGAAUCC CUUCCAAGUA AGUAUGGGCU GCCGGUACGA GCCAAGGAGU UGCUGGUUUU 240
 UUCGGAAAGA CCAUGACCGU GGAUAACAAA CUCGUUUUCC CAACGAAGGA UUUUACCAGU 300
 UUGCAACUGU GGGAGGCGUA GCUUUUGAGC AAAAACGAAG CAUAUAAUAG CUA AACACAU 360
 15 ACCGCCGACC AAAUCUACAA AGUAGUGGUG GUUAAGGUAC AUAGUACACC AGCAAAGCCA 420
 UAGAACAUAU CCAUAAAAGC AGAAGCGGUA UCGAGGAAAC ACAUGCGAAA GGAAAAGUGC 480
 UUCCAGCAUG GCCCAUCCAG CGUGAAGAGA UGGAAAGGCA CCAAAAACAA CCGGAGAGUU 540
 20 AGAAAAACCA UCAGUGUAAA UGCUAGUGCC GAAGAGAGCA UCAAUACGGG CCAAUCCACC 600
 AGGAGAGCCA CGUACUGCAU ACGUGGCAGG UUCUAAACCA UACAUUUUUU CAUACCAAGG 660
 AGGAGAACAG GGGAAAGCCA UUUGGAUAAG AACACCAAAU AAUUAUUAU AACCAAAAGU 720
 25 UCGAGCCCAA ACUGGAAGAG UUCCAGGAGG UGCAAAGAUG AAAAGAAUAA AUGAAAUGAU 780
 AAAAGGAGCC GAAUAAUGCA UGACUCCAUA UGGAACCCAG GCCAAAAUAA CAAGGAUGCU 840
 AUGCGUGGUU UUCGAGAGAA GACUAGAAAG AUUAGAGCCA UAAAGAAUAA UUUCAAGUGU 900
 30 GGGUAAAACA CGAACCCAUA UGGGUGGACG CCAGCGUUCU GGAUAAACC UACAAGAGUA 960
 AAUAAAAAUU GCCCAGGUGA UGAUAACAAU GGCAGGAAAA AAAAUUUGGC GUGUAAAAGG 1020
 AACGGUCAAC GCAUUGGCA AAAGACAGGC AAUGCCAAAU UUCCCCAGA AUCCAGGAGA 1080
 35 UUCAUUGACA AUACAAGCAA AAUCAAUUU ACCUGCUAGA AACACAUUUU GCAAUUGUGU 1140
 CCAUGACCAU UUCGUUUUGC GUAGCAAACG AAUUGUAGGC AUAGGGUUUA AGCUUGUUUC 1200
 40 CAACUUGUAA UGGGAUGCUC GGUUACACGC AGCAAGGCGC UUUUUUAAGG UCGAAAGAGC 1260
 AGACAUUGCU UCAAAGAAUU AUCAGAGUAA AAAAGGGAAG CGUACGAAAA AAUUAUCGUA 1320
 AGGAAUUAAC CGGAAACUA AAGGAAAAAA AAGGAAUUUU UAUGAAGGAA AGAAAGUAGC 1380
 45 UAUUAAUUGC AAGUGUCAAG CACUAAAAAG UAGCGAUGUA AAUUAUUUAA AAAAAGAUGG 1440
 ACCGAUUAAC CAAUGUUCAG CUCACAGUUG CCAGCAAUCA GGGCUAUUUU UUUUUUUUUU 1500
 UUAUAAAAAU GCUAAUUAU UAUAUUAUA UUAGUUUAU AACUUGCUUU UCCUCAAAAA 1560
 50 ACCAAUUCGA GAAAGGAACU UUUGCAGAGG CAAAAAGCU U 1601

55

SEQ ID NO : 19

SEQUENCE LENGTH : 12

SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Cys Phe Thr Ser Ser Tyr Phe Pro Asp Asp Arg Arg

5

10

SEQ ID NO : 20

SEQUENCE LENGTH : 19

SEQUENCE TYPE : amino acid

STRANDEDNESS : single

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

Cys Tyr Thr Ser Ile Glu Lys Tyr Asp Ile Ser Lys Ser Asp Pro

5

10

15

Leu Ala Ala Asp

SEQ ID NO : 21

SEQUENCE LENGTH : 1553

SEQUENCE TYPE : nucleic acid

STRANDEDNESS : double

TOPOLOGY : linear

MOLECULE TYPE : Genomic DNA

SEQUENCE DESCRIPTION :

TTTACATAT ATTATTCACC CAATATCATA ACAAAAACAA ACTGAATGAT GGCATCTTCT 60

ATTTTGC GTT CCAAAATAAT ACAAAAACCG TACCAATTAT TCCACTACTA TTTTCTTCTG 120

GAGAAGGCTC CTGGTTCTAC AGTTAGTGAT TTGAATTTTG ATACAAACAT ACAAACGAGT 180
TTACGTAAAT TAAAGCATCA TCATTGGACG GTGGGAGAAA TATTCCATTA TGGGTTTTTG 240
5 GTTTCATAC TTTTTTCGT GTTGTGGTT TTCCAGCTT CATTTTTTAT AAAATTACCA 300
ATAATCTTAG CATTTGCTAC TTGTTTTTA ATACCCTTAA CATCACAATT TTTCTTCCT 360
GCCTTGCCCG TTTTCACTTG GTTGGCATT TATTTTACGT GTGCTAAAAT ACCTCAAGAA 420
10 TGGAAACCAG CTATCACAGT TAAAGTTTTA CCAGCTATGG AAACAATTTT GTACGGCGAT 480
AATTTATCAA ATGTTTTGGC AACCATCACT ACCGGAGTGT TAGATATATT GGCATGGTTA 540
CCATATGGGA TTATTCATTT CAGTTTCCCA TTTGTACTTG CTGCTATTAT ATTTTTATTT 600
15 GGGCCACCGA CGGCATTAAG ATCATTTGGA TTTGCCTTTG GTTATATGAA CTGCTTGGA 660
GTCTTGATT CAAATGGCATT CCCAGCTGCT CCTCCATGGT AAAAAACTT GCACGGATTA 720
GAACCAGCTA ATTATTCAAT GCACGGGTCT CCTGGTGGAC TTGGAAGGAT AGATAAATTG 780
20 TTAGGTGTTG ATATGTATAC CACAGGGTTT TCCAATTCAT CAATCATTTT TGGGGCATT C 840
CCATCGTTAC ATTCAGGATG TTGTATCATG GAAGTGTTAT TTTTGTGTTG GTTGTTCCTA 900
CGATTCAAGT TTGTGTGGGT TACATACGCA TCTTGGCTTT GGTGGAGCAC CATGTATTTG 960
25 ACCCATCACT ACTTTGTGCA TTTGATTGGT GGAGCCATGC TATCTTTGAC TGTTTTTGAA 1020
TTCACCAAAT ATAAATATTT GCCAAAAAAC AAAGAAGGCC TTTTCTGTCT TTGGTCATAC 1080
ACTGAAATTG AAAAAATCGA TATCCAAGAG ATTGACCCTT TATCATACAA TTATATCCCT 1140
30 GTCAACAGCA ATGATAATGA AAGCAGATTG TATACGAGAG TGTACCAAGA GCCTCAGGTT 1200
AGTCCCCCAC AGAGAGCTGA AACACCTGAA GCATTTGAGA TGTCAAATTT TTCTAGGTCT 1260
AGACAAAGCT CAAAGACTCA GGTTCATTG AGTAATCTTA CTAACAATGA TCAAGTGCCT 1320
35 GGAATTAACG AAGAGGATGA AGAAGAAGAA GGCGATGAAA TTTCGTCGAG TACTCCTTCG 1380
GTGTTTGAAG ACGAACCACA GGGTAGCACA TATGCTGCAT CCTCAGCTAC ATCAGTAGAT 1440
40 GATTTGGATT CCAAAAGAAA TTAGTAAAC AGCAGTTTCT ATTAATTTCT TTATTTCTC 1500
CTAATTAATG ATTTTATGTT CAATACCTAC ACTATCTGTT TTTAATTTCC TAC 1553

45
SEQ ID NO : 22

SEQUENCE LENGTH : 472

50
SEQUENCE TYPE : amino acid

STRANDEDNESS : single

55

TOPOLOGY : linear

MOLECULE TYPE : peptide

SEQUENCE DESCRIPTION :

	Met Met Ala Ser Ser Ile Leu Arg Ser Lys Ile Ile Gln Lys Pro	
	1	5 10 15
10	Tyr Gln Leu Phe His Tyr Tyr Phe Leu Leu Glu Lys Ala Pro Gly	
	20 25 30	
15	Ser Thr Val Ser Asp Leu Asn Phe Asp Thr Asn Ile Gln Thr Ser	
	35 40 45	
	Leu Arg Lys Leu Lys His His His Trp Thr Val Gly Glu Ile Phe	
20	50 55 60	
	His Tyr Gly Phe Leu Val Ser Ile Leu Phe Phe Val Phe Val Val	
	65 70 75	
25	Phe Pro Ala Ser Phe Phe Ile Lys Leu Pro Ile Ile Leu Ala Phe	
	80 85 90	
	Ala Thr Cys Phe Leu Ile Pro Leu Thr Ser Gln Phe Phe Leu Pro	
30	95 100 105	
	Ala Leu Pro Val Phe Thr Trp Leu Ala Leu Tyr Phe Thr Cys Ala	
	110 115 120	
35	Lys Ile Pro Gln Glu Trp Lys Pro Ala Ile Thr Val Lys Val Leu	
	125 130 135	
	Pro Ala Met Glu Thr Ile Leu Tyr Gly Asp Asn Leu Ser Asn Val	
40	140 145 150	
	Leu Ala Thr Ile Thr Thr Gly Val Leu Asp Ile Leu Ala Trp Leu	
	155 160 165	
45	Pro Tyr Gly Ile Ile His Phe Ser Phe Pro Phe Val Leu Ala Ala	
	170 175 180	
50	Ile Ile Phe Leu Phe Gly Pro Pro Thr Ala Leu Arg Ser Phe Gly	
	185 190 195	

EP 0 644 262 A2

	Phe Ala Phe Gly Tyr Met Asn Leu Leu Gly Val Leu Ile Gln Met		
	200	205	210
5	Ala Phe Pro Ala Ala Pro Pro Trp Tyr Lys Asn Leu His Gly Leu		
	215	220	225
10	Glu Pro Ala Asn Tyr Ser Met His Gly Ser Pro Gly Gly Leu Gly		
	230	235	240
	Arg Ile Asp Lys Leu Leu Gly Val Asp Met Tyr Thr Thr Gly Phe		
	245	250	255
15	Ser Asn Ser Ser Ile Ile Phe Gly Ala Phe Pro Ser Leu His Ser		
	260	265	270
20	Gly Cys Cys Ile Met Glu Val Leu Phe Leu Cys Trp Leu Phe Pro		
	275	280	285
	Arg Phe Lys Phe Val Trp Val Thr Tyr Ala Ser Trp Leu Trp Trp		
25	290	295	300
	Ser Thr Met Tyr Leu Thr His His Tyr Phe Val Asp Leu Ile Gly		
	305	310	315
30	Gly Ala Met Leu Ser Leu Thr Val Phe Glu Phe Thr Lys Tyr Lys		
	320	325	330
	Tyr Leu Pro Lys Asn Lys Glu Gly Leu Phe Cys Arg Trp Ser Tyr		
35	335	340	345
	Thr Glu Ile Glu Lys Ile Asp Ile Gln Glu Ile Asp Pro Leu Ser		
	350	355	360
40	Tyr Asn Tyr Ile Pro Val Asn Ser Asn Asp Asn Glu Ser Arg Leu		
	365	370	375
	Tyr Thr Arg Val Tyr Gln Glu Pro Gln Val Ser Pro Pro Gln Arg		
45	380	385	390
	Ala Glu Thr Pro Glu Ala Phe Glu Met Ser Asn Phe Ser Arg Ser		
	395	400	405
50	Arg Gln Ser Ser Lys Thr Gln Val Pro Leu Ser Asn Leu Thr Asn		

55

410 415 420
 Asn Asp Gln Val Pro Gly Ile Asn Glu Glu Asp Glu Glu Glu Glu
 5
 425 430 435
 Gly Asp Glu Ile Ser Ser Ser Thr Pro Ser Val Phe Glu Asp Glu
 10
 440 445 450
 Pro Gln Gly Ser Thr Tyr Ala Ala Ser Ser Ala Thr Ser Val Asp
 15
 455 460 465
 Asp Leu Asp Ser Lys Arg Asn
 470
 20

Claims

1. An isolated gene coding for a protein which regulates aureobasidin sensitivity.
- 25 2. An isolated gene as claimed in Claim 1 which is contained in a DNA fragment represented by a restriction enzyme map as specified in any of Fig. 1 to Fig. 3.
3. An isolated gene as claimed in Claim 1 which is hybridizable with a gene of Claim 2.
- 30 4. A process for cloning a gene of Claim 1 characterized by using a gene of Claim 2 or 3 or a part thereof as a probe.
5. A nucleic acid probe which comprises a sequence consisting of 15 or more bases and is hybridizable with a gene of Claim 1.
- 35 6. An antisense DNA of a gene which codes for a protein regulating aureobasidin sensitivity.
7. An antisense RNA of a gene which codes for a protein regulating aureobasidin sensitivity.
- 40 8. A recombinant plasmid containing a gene of Claim 1.
9. A transformant having a recombinant plasmid of Claim 8 introduced thereinto.
- 45 10. A process for producing a protein regulating aureobasidin sensitivity characterized by culturing a transformant of Claim 9 and collecting the protein regulating aureobasidin sensitivity from the culture.
11. An isolated protein regulating aureobasidin sensitivity which is encoded by a gene of Claim 1.
- 50 12. An antibody against a protein of Claim 11.
13. A process for detecting a protein regulating aureobasidin sensitivity which comprises using an antibody of Claim 12.
- 55 14. A process for detecting a gene coding for a protein regulating aureobasidin sensitivity which comprises the hybridization with the use of a nucleic acid probe of Claim 5.

15. A process for screening an antimycotic which comprises using a transformant of Claim 9 or a protein of Claim 11.

5

10

15

20

25

30

35

40

45

50

55

Fig. 1

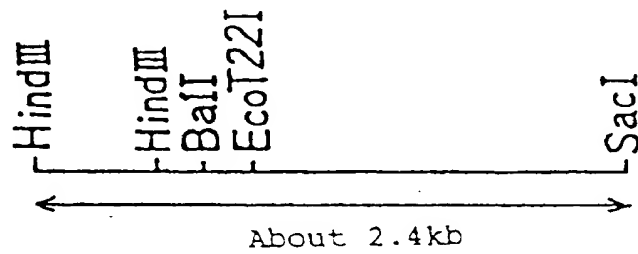


Fig. 2

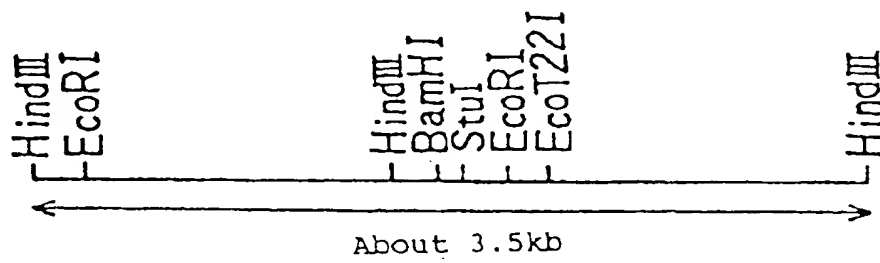


Fig. 3

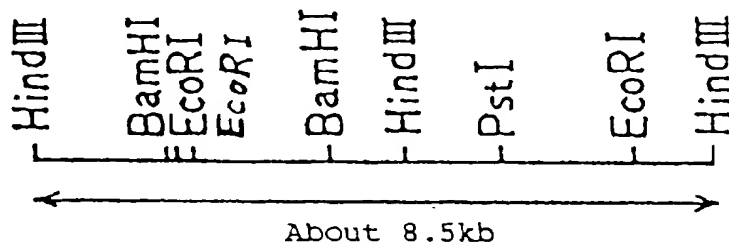


Fig. 4

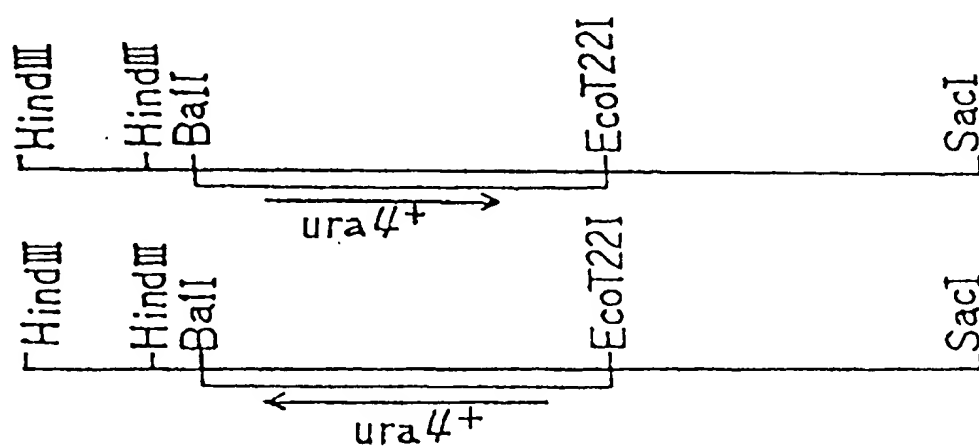


Fig. 5

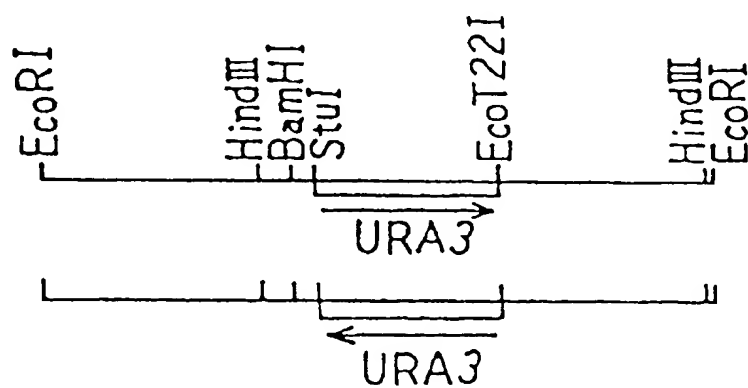


Fig. 6

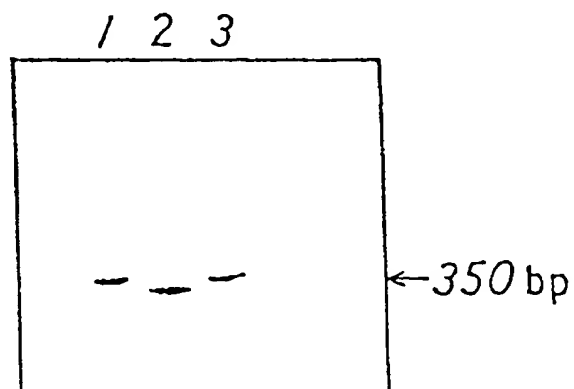


Fig. 7

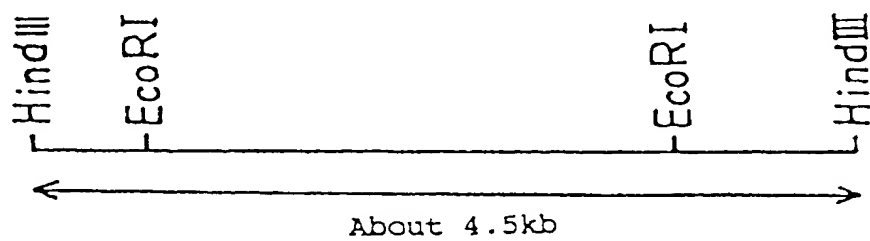


Fig. 8

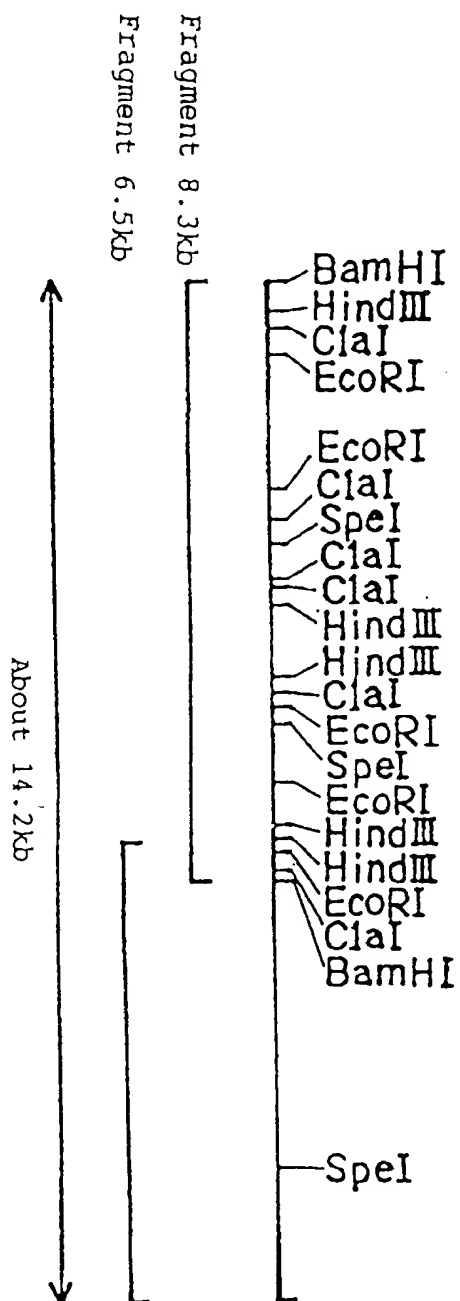


Fig. 9

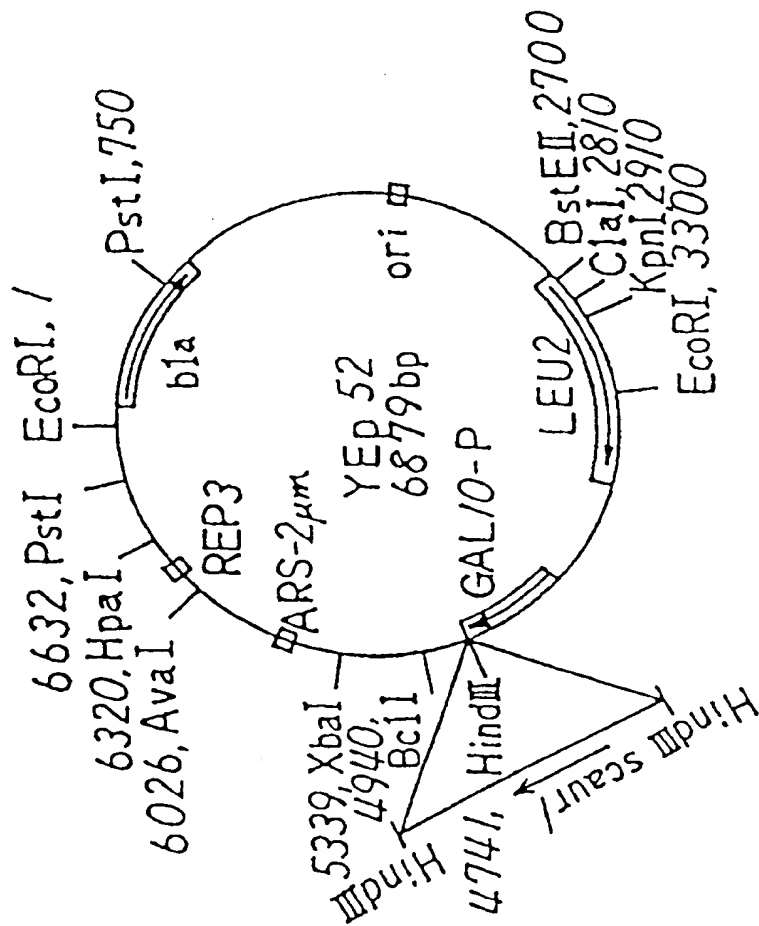


Fig. 10

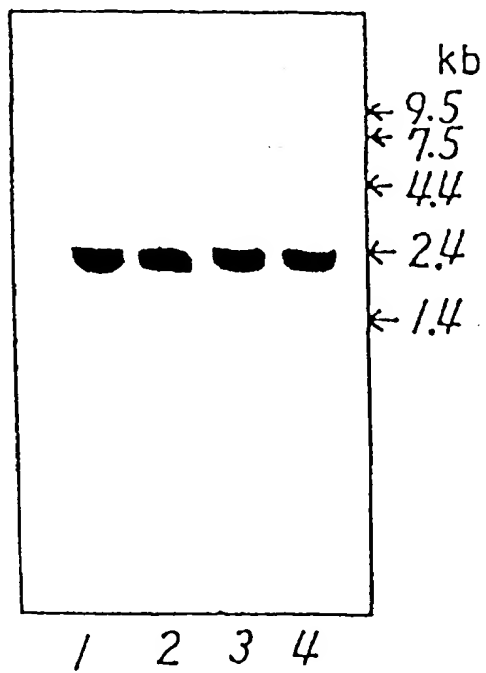


Fig. 11

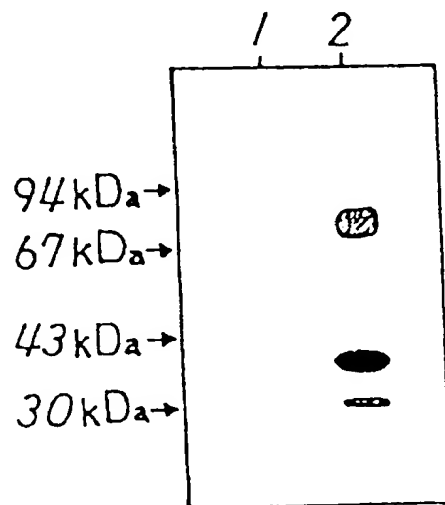
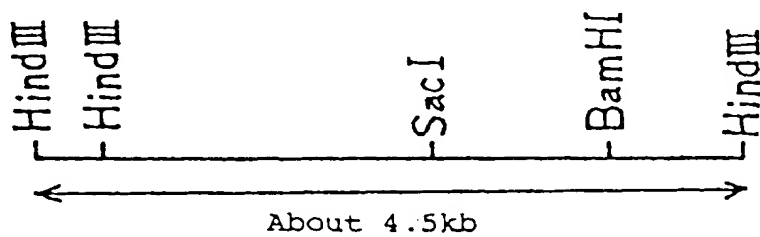


Fig. 12



(19)



Europäisches Patentamt
European Patent Office
Office européen des brevets



(11)

EP 0 644 262 A3

(12)

EUROPEAN PATENT APPLICATION

(88) Date of publication A3:
28.08.1996 Bulletin 1996/35

(51) Int. Cl.⁶: **C12N 15/31**, C07K 13/00,
C12Q 1/68, G01N 33/569,
C12N 15/11

(43) Date of publication A2:
22.03.1995 Bulletin 1995/12

(21) Application number: **94303401.7**

(22) Date of filing: **11.05.1994**

(84) Designated Contracting States:
CH DE ES FR GB IT LI

(30) Priority: **24.05.1993 JP 142523/93**
28.12.1993 JP 348893/93

(71) Applicant: **TAKARA SHUZO CO. LTD.**
Fushimi-ku, Kyoto-shi, Kyoto-fu (JP)

(72) Inventors:
• **Takashi, Okado**
Otsu-shi, Shiga-ken (JP)

• **Kazutoh, Takesako**
Otsu-shi, Shiga-ken (JP)
• **Ikunoshin, Kato**
Uji-Shi, Kyoto-fu (JP)

(74) Representative: **Wakerley, Helen Rachael**
Reddie & Grose,
16 Theobalds Road
London WC1X 8PL (GB)

(54) **Gene coding for a protein regulating aureobasidin sensitivity**

(57) An isolated gene coding for a protein regulating aureobasidin sensitivity. A process for cloning the gene with the use of the gene or a part of the same as a probe. A nucleic acid probe being hybridizable with the gene. An antisense DNA or RNA of the gene. A recombinant or transformant having the gene contained therein. An isolated protein regulating aureobasidin sensitivity and a process for producing the same by using the transformant. An antibody for the protein. A process for detecting the protein or the gene. A process for screening an antimycotic by using the protein or the transformant. This invention is useful in the diagnosis and treatment of diseases including mycoses.

EP 0 644 262 A3



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 94 30 3401

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.5)
A	EP-A-0 443 719 (TAKARA SHUZO CO. LTD) ---	1	C12N15/31 C07K13/00 C12Q1/68 G01N33/569 C12N15/11
A	MOL. GEN. GENET., vol. 235, 1992, pages 64-73, XP002007445 P. HAFFTER ET AL.: "Suppression of carboxy-terminal truncations of the yeast mitochondrial mRNA-specific translational activator PET122 by mutations in two new genes MRP17 and PET127" * figure 3 * -----		<div>TECHNICAL FIELDS SEARCHED (Int.Cl.5)</div> <div>C07K C12N C12Q G01N</div>
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 4 July 1996	Examiner Skelly, J
<div>CATEGORY OF CITED DOCUMENTS</div> <div> X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document </div>			

EPO FORM 1503 (01.02.1994) (P04C01)